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(54) Title: A METHOD FOR GENERATING BIRNAVIRUS FROM SYNTHETIC RNA TRANSCRIPTS

(57) Abstract

A system for the generation of live Birnavirus such as infectious bursal disease virus (IBDV), a segmented double-stranded (ds)RNA virus of the *Birnavirdae* family, using synthetic transcripts derived from cloned DNA has been developed. Independent full-length cDNA clones were constructed which contained the entire coding and non-coding regions of RNA segments A and B of IBDV, respectively. Synthetic RNAs of both segments were produced by *in vitro* transcription of linearized plasmids with T7 RNA polymerase. Transfection of Vero cells with combined plus-sense transcripts of both segments generated infectious virus as early as 36 hours post-transfection. The development of a reverse genetics system for dsRNA viruses will greatly facilitate studies of the regulation of viral gene expression pathogenesis, and design of a new generation of live and inactivated vaccines.



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A METHOD FOR GENERATING BIRNAVIRUS FROM SYNTHETIC RNA TRANSCRIPTS

Background of the Invention

Infectious bursal disease virus (IBDV), a member of the *Bimaviridae* family, is the causative agent of a highly immunosuppressive disease in young chickens (Kibenge, F.S.B., et al., *J. Gen. Virol.*, 69, 1757-1775 (1988)). Infectious bursal disease (IBD) or Gumboro disease is characterized by the destruction of lymphoid follicles in the bursa of Fabricius. In a fully susceptible chicken flock of 3-6 weeks of age the clinical disease causes severe immunosuppression, and is responsible for losses due to impaired growth, decreased feed efficiency, and death. Susceptible chickens less than 3 weeks old do not exhibit outward clinical signs of the disease but have a marked infection characterized by gross lesions of the bursa.

The virus associated with the symptoms of the disease is called infectious bursal disease virus (IBDV). IBDV is a pathogen of major economic importance to the nation and world's poultry industries. It causes severe immunodeficiency in young chickens by destruction of precursors of antibody-production B cells in the bursa of Fabricius. Immunosuppression causes increased susceptibility to other diseases, and interferes with effective vaccination against Newcastle disease, Marek's disease and infectious bronchitis disease viruses.

There are two known serotypes of IBDV. Serotype I viruses are pathogenic to chickens whereas serotype II viruses infect chickens and turkeys. The infection of turkeys is presently of unknown clinical significance.

IBDV belongs to a group of viruses called *Bimaviridae* which includes other bisegmented RNA viruses such as infectious pancreatic necrosis virus (fish), tellina virus and oyster virus (bivalve mollusks) and drosophila X virus (fruit fly). These viruses all contain high molecular weight (MW) double-stranded RNA genomes.

The capsid of the IBDV virion consists of several structural proteins.

As many as nine structural proteins have been reported but there is evidence that some of these may have a precursor-product relationship (Kibenge,

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F.S.B., et al., *J. Gen. Virol.*, 69, 1757-1775 (1988)). The designation and molecular weights of the viral proteins (VP) are as shown below.

5	Viral Protein	Molecular Weight
5 -	VP1	90 kDa
	VP2	41 kDa
	VP3	32 kDa
	VP4	28 kDa
10	VP5	17 kDa

Two segments of double-stranded RNA were identified in the genome of IBDV. The IBDV genome consists of two segments of double-stranded (ds)RNA that vary between 2827 (segment B) to 3261 (segment A) nucleotide base pairs (Mundt, E. et al., Virology, 209, 10-18 (1995)). The larger segment A encodes a polyprotein which is cleaved by autoproteolysis to form mature viral proteins VP2, VP3 and VP4 (Hudson, P.J. et al., Nucleic Acids Res., 14, 5001-5012 (1986)). VP2 and VP3 are the major structural proteins of the virion. VP2 is the major host-protective immunogen of IBDV, and contains the antigenic regions responsible for the induction of neutralizing antibodies (Azad, et al., Virology, 161, 145-152 (1987)). A second open reading frame (ORF), preceding and partially overlapping the polyprotein gene, encodes a protein (VP5) of unknown function that is present in IBDV-infected cells (Mundt, E., et al., J. Gen. Virol., 76, 437-443, (1995)). The smaller segment B encodes VP1, a 90-kDa multifunctional protein with polymerase and capping enzyme activities (Spies, U., et al., Virus Res., 8, 127-140 (1987): Spies, U., et al., J. Gen. Virol., 71, 977-981 (1990)).

It has been demonstrated that the VP2 protein is the major host protective immunogen of IBDV, and that it contains the antigenic region responsible for the induction of neutralizing antibodies. The region containing the neutralization site has been shown to be highly conformation-dependent. The VP3 protein has been considered to be a group-specific antigen because

it is recognized by monoclonal antibodies directed against it from strains of both serotype I and II viruses. The VP4 protein appears to be a virus-coded protease that is involved in the processing of a precursor polyprotein of the VP2, VP3 and VP4 proteins.

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Although the nucleotide sequences for genome segments A and B of various IBDV strains have been published, it was only recently that the complete 5'- and 3'-noncoding sequences of both segments were determined. The 5'-noncoding region of IBDV segments A and B contain a consensus sequence of 32 nucleotides, whereas the 3'-noncoding terminal sequences of both segments are unrelated, but conserved among IBDV strains of the same serotype (Mundt, E. et al., *Virology*, 209, 10-18 (1995)). These terminii might contain sequences important in packaging and in the regulation of IBDV gene expression, as demonstrated for other dsRNA containing viruses such as mammalian and plant reoviruses, and rotaviruses (Anzola, et al., *Proc. Natl. Acad. Sci. USA*, 84, 8301-8305 (1987); Zou, S., et al., *Virology*, 186, 377-388 (1992); Gorziglia, M.I., et al., *Proc. Natl. Acad. Sci. USA*, 89, 5784-5788 (1992)).

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In recent years, a number of infectious animal RNA viruses have been generated from cloned cDNA using transcripts produced by DNA-dependent RNA polymerase (Boyer, J.C., et al., *Virology*, 198, 415-426 (1994)). For example poliovirus, a plus-stranded RNA virus; influenza virus, a segmented negative-stranded RNA virus; rabies virus, a non-segmented negative-stranded RNA virus; all were recovered from cloned cDNAs of their respective genomes (van der Werf, S., et al., *Proc. Natl. Acad. Sci. USA*, 83, 2330-2334 (1986); Enami, M., et al., *Proc. Natl. Acad. Sci. USA*, 87, 3802-3805 (1990); Schnell, M.J., et al., *EMBO J.*, 13, 4195-4205 (1994)). For reovirus, it was shown that transfection of cells with a combination of SSRNA, dsRNA and *in vitro* translated reovirus products generated infectious reovirus when complemented with a helper virus from a different serotype (Roner, M.R., et al., *Virology*, 179, 845-852 (1990)). However, to date, there has been no report of a recovered infectious virus of segmented dsRNA genome from synthetic RNAs only.

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Summary of the Invention

This invention relates to the infectious bursal disease virus (IBDV) that is associated with Gumboro disease of young chickens. More particularly, this invention relates to a system for the generation of infectious bursal disease virus (IBDV) using synthetic transcripts derived from cloned cDNA. The present invention will facilitate studies of the regulation of viral gene expression, pathogenesis and design of a new generation of live and inactivated vaccines.

Detailed Description of the Invention

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In an effort to develop a reverse genetics system for IBDV, three independent full-length cDNA clones which contain segment A of serotype I strain D78 or serotype II strain 23/82 and segment B of the serotype I strain P2, respectively, were constructed. Synthetic RNAs of segments A and B were produced by *in vitro* transcription reaction on linearized plasmids with T7 RNA polymerase. Transcripts of these segments, either untreated or treated with DNase or RNase, were evaluated for the generation of infectious virus by transfection of Vero cells.

The present inventors have demonstrated that synthetic transcripts derived from cloned DNA corresponding to the entire genome of a segmented dsRNA animal virus can give rise to a replicating virus. The recovery of infectious virus after transfecting cells with synthetic plus-sense RNAs derived from cloned cDNA of a virus with a dsRNA genome (IBDV) completes the quest of generating reverse infectious systems for RNA viruses. A number of investigators have generated infectious animal RNA viruses from cloned cDNA (Boyer, J.C., et al., *Virology*, 198, 415-426 (1994)). Van der Werf et al. were first to generate poliovirus, a plus-stranded RNA virus, using synthetic RNA produced by T7 RNA polymerase on cloned cDNA template (van der Werf, S., et al., *Proc. Natl. Acad. Sci. USA*, 83, 2330-2334 (1986)). later, Enami et al. rescued influenza virus, a segmented negative-stranded RNA virus (Enami, M., et al., *Proc. Natl. Acad. Sci. USA*, 87, 3802-3805 (1990)); and Schnell et al. generated rabies virus, a non-segmented negative-stranded RNA virus, from cloned cDNAs of their respective genomes (Schnell, M.J., et

al., *EMBO J.*, 13, 4195-4205 (1994)). Roner et al. developed an infectious system for a segmented dsRNA reovirus by transfecting cells with a combination of synthetic ssRNA, dsRNA, *in vitro* translated reovirus products, and complemented with a helper virus of different serotype (Roner, M.R., et al., *Virology*, 179, 845-852 (1990)). The resulting virus was discriminated from the helper virus by plaque assay. However, in this system the use of a helper virus was necessary. In contrast, the presently described reverse genetics system of IBDV does not require a helper virus or other viral proteins. Transfection of cells with plus-sense RNAs of both segments was sufficient to generate infectious virus (IBDV). The fate of the additional one or four nucleotides, respectively, transcribed at the 3'-end of segment A was not determined. However, this did not prevent the replication of the viral dsRNA. Similar effects were observed for plus-stranded RNA viruses by different investigators (Boyer, J.C., et al., *Virology*, 198, 415-426 (1994)).

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Transfection of plus-sense RNAs of both segments into the same cell was necessary for the successful recovery of IBDV. Transfected RNAs of both segments had to be translated by the cellular translation machinery. The polyprotein of segment A was presumably processed into VP2, VP3 and VP4 proteins which form the viral capsid. The translated protein VP1 of segment B probably acted as a RNA-dependent RNA polymerase and transcribed minus-strands from synthetic plus-strands of both segments, and the reaction products formed dsRNA. Recently, Dobos reported that in vitro transcription by the virion RNA-dependent RNA polymerase of infectious pancreatic necrosis virus (IPNV), a prototype virus of the Birnaviridae family, is primed by VP1 and then proceeds via an asymmetric, semiconservative, stranddisplacement mechanism to synthesize only plus strands during replication of the viral genome (Dobos, P., Virology, 208, 10-25 (1995)). The present system shows that synthesis of minus-strands proceeds on the plus-strands. Whether the resulting transcribed minus-strand RNA serves as a template for the transcription of plus-strands or not remains the subject of further investigation.

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To prove that the infectious IBDV contained in the supernatants of transfected cells was indeed derived from the synthetic transcripts, an artificial chimera was generated containing segment A of a serotype II strain and segment B of a serotype I strain. Sequence analysis verified this genome combination. The results also indicate that the terminal sequence motifs described by Mundt and Müller are probably responsible for replication, sorting and packaging of the viral genome (Mundt, E. et al., *Virology*, 209, 10-18 (1995)). Presence of serotype-specific terminal sequences obviously does not prevent proper replication of serotype II A segment by the action of the RNA-dependent RNA polymerase VP1 of the serotype I segment B. The ability to create recombinant viruses will greatly help in analyzing the precise function of serotype-specific and serotype-common terminal sequences.

The recovery of infectious IBDV demonstrates that only the plus-strand RNAs of both segments are sufficient to initiate replication of dsRNA. Thus, the results are in agreement with the general features of reovirus and rotavirus replication where the plus-strand RNAs serve as a template for the synthesis of progeny minus-strands to yield dsRNA (Schonberg, M., et al., *Proc. Natl. Acad. Sci.* Patton, J.T., *Virus Res.*, 6, 217-233 (1986); Chen, D., et al., *J. Virol.*, 68, 7030-7039 (1994)). However, the semiconservative, strand displacement mechanisms proposed by Spies et al. and Dobos could not be excluded (Spies, U., et al., *Virus Res.*, 8, 127-140 (1987); Dobos, P., *Virology*, 208, 10-25 (1995)). The development of a reverse genetics system for IBDV will greatly facilitate future studies of gene expression, pathogenesis, and help in the design of new generations of live and inactivated IBDV vaccines.

As used in the present application, the term "synthetic" as applied to nucleic acids indicates that it is a man made nucleic acid in contrast to a naturally occurring nucleic acid. The term implies no limitation as to the method of manufacture, which can be chemical or biological as long as the method of manufacture involves the intervention of man.

The term "cDNA" is intended to encompass any cDNA containing segments A and B and the 5' and 3' noncoding regions of segments A and B.

The term "infectious" as applied to viruses indicates that the virus has the ability to reproduce. The virus can be pathogenic or nonpathogentic and still be infectious.

The present invention provides a system for the generation of infectious bursal disease virus using synthetic RNA transcripts. This system can be used to study the regulation of viral gene expression, pathogenesis, and for the design of a new generation of live and inactivated IBDV vaccines.

The present invention provides a recombinant vector containing at least one copy of the cDNA according to the present invention. The recombinant vector may also comprise other necessary sequences such as expression control sequences, markers, amplifying genes, signal sequences, promoters, and the like, as is known in the art. Useful vectors for this purpose are plasmids, and viruses such as baculoviruses, herpes virus (HVT) and pox viruses, e.g., fowl pox virus, and the like.

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Also provided herein is a host cell transformed with the recombinant vector of the present invention or a host cell transfected with the synthetic RNA of the present invention. The host cell may be a eukaryotic or a prokaryotic host cell. Suitable examples are *E. coli*, insect cell lines such as Sf-9, chicken embryo fibroblast (CEF) cells, chicken embryo kidney (CEK) cells, African green monkey Vero cells and the like.

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Also part of this invention is an IBDV poultry vaccine comprising a poultry protecting amount of a recombinantly produced virus or portion of a virus, wherein the virus is inactivated or modified such that it is no longer virulent.

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The virus can be inactivated by chemical or physical means. Chemical inactivation can be achieved by treating the virus with, for example, enzymes, formaldehyde, β -propiolactone, ethylene-imine or a derivative thereof, an organic solvent (e.g. halogenated hydrocarbon) and or a detergent. If necessary, the inactivating substance can be neutralized after the virus has been inactivated. Physical inactivation can be carried out by subjecting the viruses to radiation such as UV light, X-radiation, or γ -radiation.

The virus can be attenuated by known methods including serial passage, deleting sequences of nucleic acids and site directed mutagenesis either before or after production of the infectious virus to produce a virus which retains sufficient antigenicity but which has reduced virulence.

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Physiologically acceptable carriers for vaccination of poultry are known in the art and need not be further described herein. In addition to being physiologically acceptable to the poultry the carrier must not interfere with the immunological response elicited by the vaccine and/or with the expression of its polypeptide product.

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Other additives, such as adjuvants and stabilizers, among others, may also be contained in the vaccine in amounts known in the art. Preferably, adjuvants such as aluminum hydroxide, aluminum phosphate, plant and animal oils, and the like, are administered with the vaccine in amounts sufficient to enhance the immune response to the IBDV. The amount of adjuvant added to the vaccine will vary depending on the nature of the adjuvant, generally ranging from about 0.1 to about 100 times the weight of the IBDV, preferably from about 1 to about 10 times the weight of the IBDV.

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The vaccine of the present invention may also contain various stabilizers. Any suitable stabilizer can be used including carbohydrates such as sorbitol, mannitol, starch, sucrose, dextrin, or glucose; proteins such as albumin or casein; and buffers such as alkaline metal phosphate and the like. A stabilizer is particularly advantageous when a dry vaccine preparation is prepared by lyophilization.

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The vaccine can be administered by any suitable known method of inoculating poultry including nasally, ophthalmically, by injection, in drinking water, in the feed, by exposure, and the like. Preferably, the vaccine is administered by mass administration techniques such as by placing the vaccine in drinking water or by spraying the animals' environment. When administered by injection, the vaccines are preferably administered parenterally. Parenteral administration as used herein means administration by intravenous, subcutaneous, intramuscular, or intraperitoneal injection.

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The vaccine of the present invention is administered to poultry to prevent IBD anytime before or after hatching. Preferably, the vaccine is administered prior to the time of birth and after the animal is about 6 weeks of age. Poultry is defined to include but not be limited to chickens, roosters, hens, broilers, roasters, breeders, layers, turkeys and ducks.

The vaccine may be provided in a sterile container in unit form or in other amounts. It is preferably stored frozen, below -20°C, and more preferably below -70°C. It is thawed prior to use, and may be refrozen immediately thereafter. For administration to poultry the recombinantly produced virus may be suspended in a carrier in an amount of about 10⁴ to 10⁷ pfu/ml, and more preferably about 10⁵ to 10⁶ pfu/ml in a carrier such as a saline solution. The inactivated vaccine may contain the antigenic equivalent of 10⁴ to 10⁷ pfu/ml suspended in a carrier. Other carriers may also be utilized as is known in the art. Examples of pharmaceutically acceptable carriers are diluents and inert pharmaceutical carriers known in the art. Preferably, the carrier or diluent is one compatible with the administration of the vaccine by mass administration techniques. However, the carrier or diluent may also be compatible with other administration methods such as injection, eye drops, nose drops, and the like.

The invention also can be used to produce combination vaccines with the IBDV material. The IBDV material can be combined with antigen material of Newcastle Disease Virus Infectious Bronchitis virus, Reo virus, Adeno virus and/or the Marek virus.

The foregoing embodiments of the present invention are further described in the following Examples. However, the present invention is not limited by the Examples, and variations will be apparent to those skilled in the art without departing from the scope of the present invention.

Brief Description of the Drawings

Figure 1 is a schematic diagram of cDNA constructs used for synthesis of plus-sense ssRNAs of IBDV with T7 RNA polymerase. Construct pUC19FLAD78 contains the cDNA of segment A of IBDV strain D78 and the recombinant plasmid pUC18FLA23 contains the full-length cDNA of segment

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A of IBDV strain 23/82. Segment A of IBDV encodes the polyprotein (VP2-VP4-VP3), and the recently identified VP5 protein. Plasmid pUC18FLBP2 contains the cDNA of segment B of strain P2 which encodes the RNA-dependent RNA polymerase (VP1). Virus specific sequences are underlined and the T7 promoter sequences are italicized. Restriction sites are shown in boldface and identified. The cleavage sites of the linearized plasmids are shown by vertical arrows and the transcription directions are marked by horizontal arrows.

Figure 2 shows an agarose gel analysis of the transcription reaction products that were used for transfection of Vero cells. Synthetic RNAs transcribed *in vitro* using T7 RNA polymerase and linearized plasmids pUC19FLAD78 (lanes 2, 4 and 6) containing the cDNA of segment A of IBDV strain D78, and pUC18FLBP2 (lanes 1, 3 and 5) containing the cDNA of segment B of strain P2, respectively. After transcription, the reaction mixtures were either treated with DNase (lanes 1 and 2), RNase (lanes 3 and 4) or left untreated (lanes 5 and 6). Two µl of the reaction products were analyzed on 1% agarose gel. Lambda DNA, digested with *Hind* III/EcoR I, was used as markers (lane M).

Figure 3 shows a comparison of nucleotide sequences of cloned RT-PCR fragments from segments A and B of the chimeric IBDV strain 23A/P2B (bold-typed) with known sequences of segments A and B of serotype II strain 23/82 and serotype I strain P2, respectively. Nucleotide identities are marked by a colon.

Figure 4 shows the DNA sequence of pUC18FLA23.

Figure 5 shows the DNA sequence of pUC19FLAD78.

Figure 6 shows the DNA sequence of pUC18FLBP2.

EXAMPLES

Viruses and Cells. Two serotype I strains of IBDV, the attenuated P2 strain from Germany and the vaccine strain D78 (Intervet International), and one serotype II strain, the apathogenic 23/82 strain, were propagated in chicken embryo cells (CEC) and purified (Mundt, E. et al., Virology, 209, 10-18 (1995); Vakharia, V.N., et al., Virus Res., 31, 265-273 (1994)). Vero cells

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were grown in M199 medium supplemented with 5% fetal calf serum (FCS) and used for transfection experiments. Further propagation of the recovered virus and immunofluorescence studies were carried out in Vero cells (Mundt, E., et al., *J. Gen. Virol.*, 76, 437-443, (1995)). For plaque assay, monolayers of secondary CEC were prepared and used (Müller, H., et al., *Virus Res.*, 4, 297-309 (1986)).

Construction of Full-Length cDNA Clones of IBDV genome. Fulllength cDNA clones of IBDV segments A and B were independently prepared. The cDNA clones containing the entire coding region of the RNA segment A of strain D78 were prepared using standard cloning procedures and methods (Vakharia, V.N., et al., Virus Res., 31, 265-273 (1994)). By comparing the D78 terminal sequences with recently published terminal sequences of other IBDV strains (Mundt, E. et al., Virology, 209, 10-18 (1995)), it was observed that D78 cDNA clones lacked the conserved first 17 and last 10 nucleotides at the 5'- and 3'-ends, respectively. Therefore, to construct a full-length cDNA clone of segment A, two primer pairs (A5'-D78, A5-IPD78 and A3'-IPD78) were synthesized and used for PCR amplification (Table 1). The DNA segments were amplified according to the protocol of the supplier (New England Biolabs) using "Deep Vent Polymerase" (high fidelity thermophilic DNA polymerase). Amplified fragments were cloned into the EcoR I site of a pCRII vector (Invitrogen Corp.) to obtain plasmids pCRD78A5' and pCRD78A3', respectively. Each plasmid was digested with EcoR I and Sal I and the resultant fragments were ligated into EcoR I digested pUC19 to obtain plasmid pUC19FLAD78 (SEQ ID NOS:27 AND 29) which now contains a full-length cDNA copy of segment A encoding all the structural proteins (VP2, VP4 and VP3, SEQ ID NO:30) as well as the non-structural VP5 protein (SEQ ID NO:28) (Fig. 1).

Two primer pairs (A5'-23, A5IP23 and A3'-23, A3-IP23; see Table 1) were used for reverse transcription (RT) of viral genomic dsRNA of strain 23/82 using "SuperScript RT II" (RNA directed DNA polymerase with reduced RNase H activity, GIBCO/BRL). The RT reaction products were purified by phenol/chloroform extraction and ethanol precipitation. To obtain two cDNA

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fragments bounded by primer pairs A5'-23, A5-IP23 and A3'-23, A3-IP23, respectively, RT reaction products were amplified by PCR using "Deep Vent polymerase". Both RT and PCR were carried out according to the supplier's protocol. Resulting PCR fragments were blunt-end ligated into *Sma* I cleaved pUC18 vector to obtain pUC23A5' and pUC23A3'. The 3'-end of segment A contained in plasmid pUC23A3' was ligated into the *Hind* III-*Bst*B I cleaved plasmid pUC23A5' to establish the full-length cDNA of segment A of strain 23/82. The resulting plasmid was termed pUC18FLA23 (SEQ ID NOS: 31 AND 33)(Fig. 1) and encodes structural proteins VP2, VP3 and VP4 (SEQ ID NO: 32) and non-structural protein VP5 (SEQ ID NO: 34)

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To obtain cDNA clones of segment B of P2 strain, two primer pairs (B5'-P2, B5-IPP2 and B3'-P2, B3-IPP2) were designed according to the published sequences and used for RT-PCR amplification (see Table 1). Using genomic dsRNA as template, cDNA fragments were synthesized and amplified according to the supplier's protocol (Perkin-Elmer Cetus). Amplified fragments were blunt-end ligated into Sma I cleaved pBS vector (Stratagene) to obtain clones pBSP2B5' and pBSP2B3'. To construct a full-length clone of segment B, the 5'-end fragment of plasmid pBSP2B5' was first subcloned between EcoR I and Pst I sites of pUC18 vector to obtain pUCP2B5'. Then the 3'-end fragment of plasmid pBSP2B3' was inserted between the unique Bal II and Pst I sites of plasmid pUCP2B5' to obtain a full-length plasmid pUC18FLBP2 (SEQ ID NO:25) which encodes the VP1 protein (SEQ ID NO: 26) (Fig. 1). Plasmids pUC18FLBP2, pUC18FLA23 and pUC19FLAD78 were completely sequenced by using the "Sequenase" DNA sequencing system (U.S. Biochem.), and the sequence data were analyzed using either "DNASIS" (Pharmacia) or "PC/Gene" (Intelligenetics) software. The integrity of the full-length constructs was tested by in vitro transcription and translation coupled reticulocyte lysate system using T7 RNA polymerase (Promega).

Transcription and Transfection of Synthetic RNAs. Plasmids pUC19FLAD78, pUC18FLA23 and pUC18FLBP2 were digested with *BsrGI*, *NsiI* and *PstI* enzymes (see Fig. 1), respectively, and used as templates for *in vitro* transcription with T7 RNA polymerase (Promega). Briefly, restriction

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enzyme cleavage assays were adjusted to 0.5% SDS and incubated with proteinase K (0.5 mg/ml) for 1 hour at 37°C. The linearized DNA templates (~3 µg) were recovered after ethanol precipitation, and were added separately to a transcription reaction mixture (50 µl) containing 40 mM Tris-HCl (pH 7.9), 10 mM NaCl, 6 mM MgCl₂, 2 mM spermidine, 0.5 mM ATP, CTP and UTP each, 0.1 mM GTP, 0.25 mM cap analog [m7G(5') PPP(5') G], 120 units of "RNasin" (ribonuclease inhibitor), 150 units T7 RNA polymerase (Promega), and incubated at 37°C for 1 hour. Synthetic RNA transcripts were purified by phenol/chloroform extraction and ethanol precipitation. As controls, the transcription products were treated with either DNase or RNase (Promega) before the purification step.

Vero cells were grown to 80% confluence in 60 mm dishes and washed once with phosphate-buffered saline (PBS). Three ml of "OPTI-MEM I" (reduced serum medium containing HEPES buffer, sodium bicarbonate, hypoxanthine, thymidine, sodium pyruvate, L-glutamine, trace elements, growth factors and phenol red; from GIBCO/BRL) were added to the monolayers, and the cells were incubated at 37°C for 1 hour in a CO₂ incubator. Simultaneously, 0.15 ml of "OPTI-MEM I" was incubated with 1.25 (N-[1-(2,3-dioleyloxy)propyl]-N,N,N-"Lipofectin" reagent μg dioleoylphosphatidylethanolamine, chloride and trimethylammonium GIBCO/BRL) for 45 min. in a polystyrene tube at room temperature. Synthetic RNA transcripts of both segments, resuspended in 0.15 ml of diethyl pyrocarbonate-treated water, were added to the OPTI-MEM-Lipofectinmixture, mixed gently, and incubated on ice for 5 min. After removing the "OPTI-MEM" from the monolayers in 60 mm dishes and replacing with fresh 1.5 ml of "OPTI-MEM", the nucleic acid containing mixture was added dropwise to the Vero cells and swirled gently. After 2 hours of incubation at 37°C, the mixture was replaced with M199 medium [CaCl₂ (annhydrous), Fe(NO₃)₃ 9H₂0, KCl, MgSO₄ (anhydrous), NaCl, NaH₂PO₄H₂O, NaHCO₃, L-Alanine, L-Arginine HCl, L-Aspartic acid, L-Cysteine HCl H2O, L-Cysteine 2HCl, L-Glutamic acid, L-Glutamine, Glycine, L-Histidine HCL H₂O, L-Hydroxyproline, L-Isoleucine, L-Leucine, L-Lysine HCI, L-Methionine, L-Phenylalanine, L-

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Proline, L-Serine, L-Threonine, L-Tryptophan, L-Tyrosine 2Na 2H₂O, L-Valine, Alpha tocopherol PO₄ Na₂, Ascorbic Acid, Biotin, Calciferol, D-Calcium pantothenate, Choline chloride, Folic acid, I-Inositol, Menandione NaHSO₃ 3H₂O, Niacin, Nicotinamide, Para-aminobenzoic acid, Pyridoxine HCl, Riboflavin, Thiamine HCl, Vitamin A Acetate, Adenine SO₄, Adenylic Acid, ATP, Na₂, Cholesterol, 2-Deoxy-D-Ribose, D-Glucose, Glutathione, Guanine HCl, Hypoxanthine Na, Phenol Red Na, Ribose, Sodium Acetate (anhydrous), Thymine, Tween 80, Uracil, and Xanthine Na; from Mediatech, Inc.] containing 5% FCS (without rinsing cells) and the cells were further incubated at 37°C for desired time intervals.

Identification of Generated IBDV. CEC were infected with filtered (0.2 μm) supernatant from Vero cells transfected with transcripts of pUC18FLA23 and pUC18FLP2B. 16 hours post-infection, the whole cell nucleic acids were isolated (Mundt, E. et al., *Virology*, 209, 10-18 (1995)). Primers were designed according to the published sequences and RT-PCR fragments were amplified, cloned and sequenced (Mundt, E. et al., *Virology*, 209, 10-18 (1995)). Sequence data were analyzed by using "DNASIS" software.

Immunofluorescence. Vero cells, grown on cover slips to 80% confluence, were infected with the supernatants derived from transfected Vero cells (after freeze-thawing) and incubated at 37°C for two days. The cells were then washed, fixed with acetone and treated with polyclonal rabbit anti-IBDV serum. After washing, the cells were treated with fluorescein labeled goat-anti-rabbit antibody (Kirkegaard & Perry Lab.) and examined by fluorescence microscope.

Plaque Assay. Monolayers of secondary CEC, grown in 60 mm dishes, were inoculated with the supernatants derived from transfected Vero cells. After 1 hour of infection, the cells were washed once with PBS and overlayed with 0.8% Agar noble (Difco) containing 10% tryptose phosphate broth, 2% FCS, 0.112% NaHCO₃, 10³ units penicillin, 10³ μg/ml streptomycin, 0.25 μg/ml fungizone, 0.005% neutral red, 0.0015% phenol red. The cells

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were incubated at 37°C for 2 to 3 days until plaques could be observed and counted (Müller, H., et al., *Virus Res.*, 4, 297-309 (1986)).

Construction of Full-Length cDNA clones of IBDV Genome. To develop a reverse genetics system for the dsRNA virus IBDV, two independent cDNA clones were constructed that contain segment A of strain D78 and segment B of strain P2 (Fig. 1). Each plasmid encoded either the precursor of structural proteins (VP2, VP4, VP3) and VP5 or only VP1 protein (RNA-dependent RNA polymerase). Plasmid pUC18FLBP2 upon digestion with Pst I and transcription in vitro by T7 RNA polymerase, would yield RNA containing the correct 5'- and 3'-ends. Whereas, upon digestion with BsrG I and transcription, plasmid pUC19FLAD78 would yield RNA containing the correct 5'-end but with additional four nucleotides at the 3'end. Coupled transcription and translation of the above plasmids in a rabbit reticulocyte system yielded protein products that were correctly processed and comigrated with the marker IBDV proteins after fractionating on SDS-polyacrylamide gel and autoradiography (data not shown).

Plus-sense transcripts of IBDV segment A and B were synthesized separately in vitro with T7 RNA polymerase using linearized full-length cDNA plasmids as templates (see Fig. 2). Although two species of RNA transcripts were observed for segment B on a neutral gel (lanes 1 and 5), fractionation of these samples on a denaturing gel yielded only one transcript-specific band (data not shown). In order to show that plus-sense RNA transcripts of both segments are needed for the generation of infectious virus, the transcription mixtures were incubated with different nucleases, as shown in Fig. 2. Synthetic RNAs recovered after treating the transcription products with DNase (lanes 1+2), RNase (lanes 3+4) or without treatment (lanes 5+6), were used for the transfection of Vero cells. As mock control, Lipofectin alone was used. Five days post-transfection, cytopathic effect (CPE) was only visible in Vero cells transfected with combined transcripts of untreated or DNase-treated transcription products, but not with RNase-treated transcription mixtures or

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mock-transfected control. In addition, no CPE was detected when Vero cells were transfected with RNA of only segment A or B (data not shown). These results demonstrate that replication of IBDV ensued after transfection of Vero cells with plus-sense ssRNAs of both segments of IBDV. To verify that the agent causing the CPE in Vero cells was indeed IBDV, transfected Vero cells were freeze-thawed, and supernatants were clarified by centrifugation, and used to infect CEC or Vero cells. CEC infected with the supernatants derived from Vero transfected cells of untreated or DNase-treated transcription mixtures produced CPE in one day post-inoculation (Table 2). However, no CPE could be detected even after five days in CEC, with the supernatants from transfected Vero cells of RNase-treated transcription mixtures, untreated segment A or B transcription mixtures and mock-transfected Vero cells. Similarly, when Vero cells on cover slips were infected with the same supernatants as described above and examined by immunofluorescence staining after 2 days, only supernatants derived from transfected Vero cells of untreated or DNAse-treated transcription mixtures gave positive immunofluorescence signal (Table 2).

Recovery of Transfectant Virus. To determine the time point for the recovery of infectious virus, Vero cells were transfected with combined RNA transcripts of segments A and B. At 4, 8, 16, 24, 36 and 48 hours post-transfection, the supernatants were examined for the presence of transfectant virus by infectivity and plaque assays, as shown in Table 3. Our results indicate that the virus could be recovered as early as 36 hours after transfection. Virus titer was 2.3 x 10² pfu/ml which appear to drop for samples obtained later than 48 hours after transfection.

Generation of a Chimeric Virus. To prove that plus-sense ssRNA of both segments of IBDV are sufficient for recovery of infectious virus, a chimeric IBDV was generated. Plasmid pUC18FLA23 containing a full-length sequence of segment A of serotype II strain was linearized by *Nsi* I digestion and ssRNA was synthesized *in vitro* using T7 RNA polymerase. The ssRNA transcript specifies the correct 5'-end but contains one additional residue at the 3'-end (Fig. 1). Vero cells were transfected with ssRNA of segment A of

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serotype II strain 23/82 and ssRNA of segment B of serotype I strain P2. Five days after transfection when CPE was evident, the supernatant was clarified (after freeze-thawing) and used to infect CEC. After a second passage in CEC, genomic RNA of the virus was analyzed by RT-PCR and sequencing of the PCR products. Primers for segment A were deigned to specifically amplify only segment A sequences derived from the serotype II strain. Primer for segment B bound to sequences of both serotypes. The amplified fragments were cloned and sequenced. The obtained segment A sequences showed a perfect match with known segment A sequences of serotype II strain 23/82, whereas segment B sequence exhibited complete homology to published segment B sequences of serotype I strain P2 (Fig. 3).

Table 1. Oligonucleotides Used for the Construction of Full Length cDNA Clones of IBDV Genomic Segments A and B.

:		
Orientation	Name	Nucleotide Number
(+)	A5'-D78	1-31
(+)	A5′-23	1-48
(-)	A3'-D78	3237-3261
(-)	A3′-23	3242-3261
(-)	A5-IPD78	1711-1730
(-)	A5-IP23	1971-1990
÷	A3-IPD78	1723-1742
(t)	A3-IP23	1883-1900
€	B5P2	1-18
(-)	B3'-P2	2807-2827
(-)	B5-IPP2	1915-1938
(+)	B3-IPP2	1839-1857
サートーナーナーコーモ		

Composition and location of the oligonucleotide primers used for cloning. T7 promoter sequences are marked with italic types, the virus specific sequences are underlined, and the restriction sites marked in boldface. Orientation of the virus specific sequence of the primer is shown for sense (+) and antisense (-). The positions where the primers bind (nucleotide number) are according to the published sequences of P2 strain (2).

Table 2. Generation of Infections IBDV From Synthetic RNAs of Segment A and B.

Material Transfected	CPE	Immunofluoroescence
ssRNA A+B, DNase-treated	+	+
ssRNA A+B, RNase-treated	-	<u>-</u> ·
ssRNA A+B, untreated	+	+
ssRNA A; untreated	<u>-</u>	-
ssRNA B, untreated	-	-
Lipofectin only	-	-

Vero cells were transfected with synthetic RNAs of segment A and B derived from transcription reactions that were either untreated or treated with DNase or RNase. After 5 days, the supernatants were collected, clarified by centrifugation, and analyzed for the presence of virus. The infectivity of the recovered virus was determined in CEC by the appearance of cytopathic effect (CPE) 1-2 days post-inoculation. The specificity of the recovered virus was determined by immunofluorescence staining of infected Vero cells with rabbit anti-IBDV serum.

Table 3. Recovery of Virus at Various Times Post-Transfection.

Time in hours post-transfection	CPE	Immunofluorescence	pfu/mi
4	-	-	0
8	_	-	0
16	-		0
24		-	0
36	+	+	2.3 × 10 ²
48	+	+	6.0 × 10 ¹

Vero cells were transfected with synthetic RNAs of segment A and B as described. The infectivity and specificity of the recovered virus was detected by CPE in CEC and immunofluorescence staining in Vero cells, respectively. Monolayers of secondary CEC were used for plaque assay after inoculating the cells with the supernatants derived from transfected Vero cells. Approximate titer of the virus was calculated as plaque forming units per ml (pfu/ml).

SEQUENCE LISTING

- (1) GENERAL INFORMATION:
 - (i) APPLICANT: VAKHARIA, Vikram N. MUNDT, Egbert
- (ii) TITLE OF INVENTION: A METHOD FOR GENERATING BIRNAVIRUS FROM SYNTHETIC RNA TRANSCRIPTS
 - (iii) NUMBER OF SEQUENCES: 34
 - (iv) CORRESPONDENCE ADDRESS:
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 - (B) STREET: 655 Fifteenth Street, N. W., Suite 330 G Street Lobby
 - (C) CITY: Washington
 - (D) STATE: DC
 - (E) COUNTRY: USA
 - (F) ZIP: 20005-5701
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
 - (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: US
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
 - (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: KITTS, Monica C.
 - (B) REGISTRATION NUMBER: 36,105
 - (C) REFERENCE/DOCKET NUMBER: P8172-6002
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 202/638-5000
 - (B) TELEFAX: 202/638-4810
- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 46 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: circular
 - (ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:	•
GAATTCGGCT TTAATACGAC TCACTATAGG ATACGATCGG TCTGAC	46
(2) INFORMATION FOR SEQ ID NO:2:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 41 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: circular 	
(ii) MOLECULE TYPE: CDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:	
AATTGGATCC GTTCGCGGGT CCCCTGTACA AAGCCGAATT C	41
(2) INFORMATION FOR SEQ ID NO:3:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 36 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: circular 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:	•
CGGCGAATTC ATGCATAGGG GACCCGCGAA CGGATC	36
(2) INFORMATION FOR SEQ ID NO:4:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 44 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: circular 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:	
STCAGACCGA TCGTATCCTA TAGTGAGTCG TATTAGAATT CTCT	44

(2) INFORMATION FOR SEQ ID NO:5:

(B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear

 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 33 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: circular 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:	
TTGCATGCCT GCAGGGGCC CCCGCAGGCG AAG	33
(2) INFORMATION FOR SEQ ID NO:6:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 31 base pairs (B) TYPE: nucleic acid	
(C) STRANDEDNESS: double	
(D) TOPOLOGY: circular	
(b) forobodi. Circuit	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:	
TCGTATCCTA TAGTGAGTCG TATTAGAATT C	31
(2) INFORMATION FOR SEQ ID NO:7:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 120 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:	
GGAAGCCTGA GTGAGTTGAC TGACTACAGC TACAACGGGC TGATGTCAGC CACTGCGAAC	60
ATCAACGACA AGATCGGGAA CGTTCTAGTT GGAGAAGGGG TGACTGTTCT CAGTCTACCG	120
(2) INFORMATION FOR SEQ ID NO:8:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 120 base pairs	

(ii) MOLECULE TYPE: DNA	-
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:	
GGAAGCCTGA GTGAGTTGAC TGACTACAGC TACAACGGGC TGATGTCAGC CACTGCGAAC	60
ATCAACGACA AGATCGGGAA CGTTCTAGTT GGAGAAGGGG TGACTGTTCT CAGTCTACC	119
(2) INFORMATION FOR SEQ ID NO:9:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 120 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:	
GGAAGCCTGA GTGAACTGAC AGATGTTAGC TACAATGGGT TGATGTCTGC AACAGCCAAC	60
ATCAACGACA AAATTGGGAA CGTCCTAGTA GGGGAAGGGG TCACCGTCCT CAGCTTACCC	120
(2) INFORMATION FOR SEQ ID NO:10:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 120 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:	
TTTTCAATAG TCCACAGGCG CGAACGAAGA TCTCAGCAGC GTTCGGCATA AAGCCTACTG	60
CTGGACAAGA CGTGGAAGAA CTCTTGATCC CCAAAGTCTG GGTGCCACCT GAGGATCCGC	120
(2) INFORMATION FOR SEQ ID NO:11:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 120 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single	

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:	
TTTTCAACAG TCCACAGGCG CGAAGCACGA TCTCAGCAGC GTTCGGCATA AAGCCTACTG	60
CTGGACAAGA CGTGGAAGAA CTCTTGATCC CTAAAGTTTG GGTGCCACCT GAGGATCCGC	120
(2) INFORMATION FOR SEQ ID NO:12:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 120 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:	
TTTTCAACAG TCCACAGGCG CGAAGCACGA TCTCAGCAGC GTTCGGCATA AAGCCTACTG	60
CTGGACAAGA CGTGGAAGAA CTCTTGATCC CTAAAGTTTG GGTGCCACCT GAGGATCCGC	120
(2) INFORMATION FOR SEQ ID NO:13: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 48 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:	
TAATACGACT CACTATAGGA TACGATCGGT CTGACCCCGG GGGAGTCA	48
(2) INFORMATION FOR SEQ ID NO:14: (i) SEQUENCE CHARACTERISTICS:	<i>.</i>
(A) LENGTH: 44 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	

(ii) MOLECULE TYPE: DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

AGA	GAATI	TCT AATACGACTC ACTATAGGAT ACGATCGGTC	TGAC	44
. (2)	INFO	ORMATION FOR SEQ ID NO:15:		
		SEQUENCE CHARACTERISTICS: (A) LENGTH: 30 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear		÷
	(ii)	MOLECULE TYPE: DNA		-
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:15:		
TGT	ACAGG	GG ACCCGCGAAC GGATCCAATT		30
(2)	INFO	RMATION FOR SEQ ID NO:16:	the second second	
	(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 36 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear		-
	(ii)	MOLECULE TYPE: DNA		
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:16:	. •	
CGGC	GAAT	TC ATGCATAGGG GACCCGCGAA CGGATC		36
(2)	INFO	RMATION FOR SEQ ID NO:17:		
	(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear		
	(ii)	MOLECULE TYPE: DNA		
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:17:		
CGTC	GACT	AC GGGATTCTGG		20
			•	

- (2) INFORMATION FOR SEQ ID NO:18:
 - (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs

TU 110071114700

		(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear			
	(ii)	MOLECULE TYPE: DNA			
	(xi)	SEQUENCE DESCRIPTION: SEQ II	NO:18:		
CAGA	GGCAG	et actccgtctg			20
(2)	Infor	RMATION FOR SEQ ID NO:19:			
	(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear			
	(ii)	MOLECULE TYPE: DNA			
	(xi)	SEQUENCE DESCRIPTION: SEQ II	NO:19:		
AGTC	GACGO	GG ATTCTTGCTT			20
(2)	INFOF	RMATION FOR SEQ ID NO:20:			
	(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear			
•	(ii)	MOLECULE TYPE: DNA			
	(xi)	SEQUENCE DESCRIPTION: SEQ I	D NO:20:		
GAAG	GTGT	GC GAGAGGAC			18
(2)	INFO	RMATION FOR SEQ ID NO:21:		·	
	(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 44 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear			
	(ii)	MOLECULE TYPE: DNA			
	(xi)	SEQUENCE DESCRIPTION: SEQ I	D NO:21:		

AGA	GAATTCT AATACGACTC ACTATAGGAT ACGATGGGTC TGAC	44
(2)	INFORMATION FOR SEQ ID NO:22:	
	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 33 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
	(ii) MOLECULE TYPE: DNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:	
CGA	CTGCTG CAGGGGCCC CCGCAGGCGA AGG	33
(2)	INFORMATION FOR SEQ ID NO:23:	
	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 24 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
	(ii) MOLECULE TYPE: DNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:	
CTTG	AGACTC TTGTTCTCTA CTCC	24
(2)	INFORMATION FOR SEQ ID NO:24:	
	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 19 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
	(ii) MOLECULE TYPE: DNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:	
ATAC	AGCAAA GATCTCGGG	19
(2)	INFORMATION FOR SEQ ID NO:25:	
	(i) SECTIFACE CHARACTERISTICS.	

(A) LENGTH: 2827 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single (D) TOPOLOGY: circular

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 112..2745

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:													
GGATACGATG GGTCTGACCC TCTGGGAGTC ACGAATTAAC GTGGCTACTA GGGGCGATAC													
CCGCCGCTGG CC	GCCACGTT AGTGG	CTCCT CTTCTTGA	TG ATTCTGCCAC C ATG AGT Met Ser 1	117									
GAC ATT TTC A Asp Ile Phe A 5	AC AGT CCA CAG Asn Ser Pro Gln	GCG CGA AGC A Ala Arg Ser T 10	CG ATC TCA GCA GCG TTC hr Ile Ser Ala Ala Phe 15	165									
GGC ATA AAG C Gly Ile Lys P 20	CT ACT GCT GGA Pro Thr Ala Gly 25	Gln Asp Val G	AA GAA CTC TTG ATC CCT lu Glu Leu Leu Ile Pro 30	213									
AAA GTT TGG G Lys Val Trp V 35	GTG CCA CCT GAG Val Pro Pro Glu 40	a Asp Pro Leu A	CC AGC CCT AGT CGA CTG la Ser Pro Ser Arg Leu 45 50	261									
GCA AAG TTC C Ala Lys Phe I	CTC AGA GAG AAC Leu Arg Glu Asr 55	GGC TAC AAA G Gly Tyr Lys V 60	TT TTG CAG CCA CGG TCT al Leu Gln Pro Arg Ser 65	309									
CTG CCC GAG F Leu Pro Glu F	AAT GAG GAG TAT Asn Glu Glu Tyr 70	G GAG ACC GAC C Glu Thr Asp G 75	AA ATA CTC CCA GAC TTA ln Ile Leu Pro Asp Leu 80	357									
GCA TGG ATG (Ala Trp Met A 85	CGA CAG ATA GAA Arg Gln Ile Gli	A GGG GCT GTT T 1 Gly Ala Val L 90	TA AAA CCC ACT CTA TCT eu Lys Pro Thr Leu Ser 95	405									
CTC CCT ATT (Leu Pro Ile (100	GGA GAT CAG GAG Gly Asp Gln Glo 10:	1 Tyr Phe Pro L	AG TAC TAC CCA ACA CAT ys Tyr Tyr Pro Thr His 110	453									
CGC CCT AGC A Arg Pro Ser I	AAG GAG AAG CCC Lys Glu Lys Pro 120	o Asn Ala Tyr P	CG CCA GAC ATC GCA CTA ro Pro Asp Ile Ala Leu 25 130	501									

CTC Leu	Lys	CAG Gln	ATG Met	ATT Ile 135	Tyr	CTG Leu	TTT	CTC Leu	CAG Gln 140	Val	CCA Pro	GAG Glu	GCC Ala	AAC Asn 145	GAG Glu	549
GGC	CTA	AAG Lys	GAT Asp 150	GAA Glu	GTA Val	ACC Thr	CTC Leu	TTG Leu 155	ACC Thr	CAA Gln	AAC Asn	ATA Ile	AGG Arg 160	qaA	AAG Lys	597
Ala	Tyr	Gly 165	Ser	Gly	Thr	Tyr	Met 170	Gly	Gln	Ala	Asn	Arg 175	Leu	Val	GCC Ala	645
ATG Met	AAG Lys 180	GAG Glu	GTC Val	GCC Ala	ACT Thr	GGA Gly 185	AGA Arg	AAC Asn	CCA Pro	AAC Asn	AAG Lys 190	GAT Asp	CCT Pro	CTA Leu	AAG Lys	693
CTT Leu 195	GGG Gly	TAC Tyr	ACT Thr	TTT Phe	GAG Glu 200	AGC Ser	ATC Ile	GCG Ala	CAG Gln	CTA Leu 205	CTT Leu	GAC Asp	ATC Ile	ACA Thr	CTA Leu 210	741
CCG Pro	GTA Val	GGC	CCA Pro	CCC Pro 215	GGT Gly	GAG Glu	GAT Asp	GAC Asp	AAG Lys 220	CCC Pro	TGG Trp	GTG Val	CCA Pro	CTC Leu 225	ACA Thr	789
AGA Arg	GTG Val	CCG Pro	TCA Ser 230	CGG Arg	ATG Met	TTG Leu	GTG Val	CTG Leu 235	ACG Thr	GGA Gly	GAC Asp	GTA Val	GAT Asp 240	GGC Gly	GAC Asp	837
TTT Phe	GAG Glu	GTT Val 245	GAA Glu	GAT Asp	TAC Tyr	CTT Leu	CCC Pro 250	AAA Lys	ATC Ile	AAC Asn	CTC Leu	AAG Lys 255	TCA Ser	TCA Ser	AGT Ser	885
						CGC Arg 265										933
						TTT Phe										981
						GGG Gly										1029
						TTA Leu										1077
						ACA Thr										1125

TCA Ser	GCT Ala 340	CCA Pro	TCC Ser	CCA Pro	ACA Thr	CAC His 345	CTC Leu	ATG Met	ATC Ile	TCT Ser	ATG Met 350	ATC Ile	ACC Thr	TGG Trp	CCC		1173
			AAC Asn														1221
TCA Ser	CTC Leu	TAC Tyr	AAA Lys	TTC Phe 375	AAC Asn	CCG Pro	TTC Phe	AGA Arg	GGA Gly 380	GGG Gly	TTG Leu	AAC Asn	AGG Arg	ATC Ile 385	GTC Val	. ·	1269
GAG Glu	TGG Trp	Ile	TTG Leu 390	GCC Ala	CCG Pro	GAA Glu	GAA Glu	CCC Pro 395	AAG Lys	GCT Ala	CTT Leu	GTA Val	TAT Tyr 400	GCG Ala	GAC Asp	٠	1317
AAC Asn	ATA Ile	TAC Tyr 405	ATT Ile	GTC Val	CAC His	TCA Ser	AAC Asn 410	ACG Thr	TGG Trp	TAC Tyr	TCA Ser	ATT Ile 415	GAC Asp	CTA Leu	GAG Glu	· .	1365
AAG Lys	GGT Gly 420	GAG Glu	GCA Ala	AAC Asn	TGC Cys	ACT Thr 425	CGC Arg	CAA Gln	CAC His	ATG Met	CAA Gln 430	GCC Ala	GCA Ala	ATG Met	TAC Tyr		1413
TAC Tyr 435	ATA Ile	CTC Leu	ACC Thr	AGA Arg	GGG Gly 440	TGG Trp	TCA Ser	GAC Asp	AAC Asn	GGC Gly 445	GAC Asp	CCA Pro	ATG Met	TTC Phe	AAT Asn 450	•	1461
CAA Gln	ACA Thr	TGG Trp	GCC Ala	ACC Thr 455	TTT Phe	GCC Ala	ATG Met	AAC Asn	ATT Ile 460	GCC Ala	CCT Pro	GCT Ala	CTA Leu	GTG Val 465	GTG Val		1509
GAC Asp	TCA Ser	TCG Ser	TGC Cys 470	CTG Leu	ATA Ile	ATG Met	AAC Asn	CTG Leu 475	CAA Gln	ATT Ile	AAG Lys	ACC Thr	TAT Tyr 480	GGT Gly	CAA Gln		1557
GGC Gly	AGC Ser	GGG Gly 485	AAT Asn	GCA Ala	GCC Ala	ACG Thr	TTC Phe 490	ATC Ile	AAC Asn	AAC Asn	CAC His	CTC Leu 495	TTG Leu	AGC Ser	ACA Thr		1605
CTA	GTG Val 500	CTT Leu	GAC Asp	CAG Gln	TGG	AAC Asn 505	CTG Leu	ATG Met	AGA Arg	CAG Gln	Pro 510	AGA Arg	CCA	GAC Asp	AGC Ser		1653
GAG Glu 515	GAG Glu	TTC Phe	AAA Lys	TCA Ser	ATT Ile 520	GAG Glu	GAC Asp	AAG Lys	CTA	GGT Gly 525	ATC Ile	AAC Asn	TTT Phe	AAG Lys	ATT Ile 530		1701
GAG Glu	AGG Arg	TCC Ser	ATT	GAT Asp 535	GAT Asp	ATC Ile	AGG Arg	GGC Gly	AAG Lys 540	CTG Leu	AGA Arg	CAG Gln	CTT	GTC Val 545	CTC Leu		1749

CTT Leu	GCA Ala	CAA Gln	CCA Pro 550	GGG Gly	TAC Tyr	CTG Leu	AGT Ser	GGG Gly 555	GGG	GTT Val	GAA Glu	CCA Pro	GAA Glu 560	Gln	TCC	1797
Ser	Pro	Thr 565	GTT Val	Glu	Leu	qaA	Leu 570	Leu	Gly	Trp	Ser	Ala 575	Thr	Tyr	Ser	1845
AAA Lys	GAT Asp 580	CTC	GGG Gly	ATC Ile	TAT	GTG Val 585	CCG	GTG Val	CTT Leu	GAC Asp	AAG Lys 590	GAA Glu	CGC Arg	CTA Leu	TTT Phe	1893
TGT Cys 595	TCT	GCT Ala	GCG Ala	TAT Tyr	CCC Pro 600	AAG Lys	GGA Gly	GTA Val	GAG Glu	AAC Asn 605	AAG Lys	AGT Ser	CTC	AAG Lys	TCC Ser 610	1941
AAA Lys	GTC Val	GGG Gly	ATC Ile	GAG Glu 615	CAG Gln	GCA Ala	TAC Tyr	AAG Lys	GTA Val 620	GTC Val	AGG Arg	TAT Tyr	GAG Glu	GCG Ala 625	TTG Leu	1989
			GGT Gly 630													2037
AAT Asn	AAC Asn	GCA Ala 645	GGC Gly	GCC Ala	GCT Ala	CGG Arg	CGG Arg 650	CAT His	CTG Leu	GAG Glu	GCC Ala	AAG Lys 655	GGG Gly	TTC Phe	CCA Pro	2085
CTC Leu	GAC Asp 660	GAG Glu	TTC Phe	CTA Leu	GCC Ala	GAG Glu 665	TGG Trp	TCT Ser	GAG Glu	CTG Leu	TCA Ser 670	GAG Glu	TTC Phe	GGT Gly	GAG Glu	2133
			GGC Gly											Ser		2181
			AAC Asn													2229
			ACT Thr 710													2277
			AGG Arg			Ala										2325
			AGC Ser													2373

GAG	AAA	CTC	CAC	AAG	TCC Ser	AAG Lvs	CCA Pro	GAC Asp	GAC	CCC	GAT	GCA Ala	GAC Asp	TGG	TTC Phe	2421
755	_				760					765					770	
GAA	AGA	TCA	GAA	ACT	CTG	TCA	GAC	CTT	CTG	GAG	AAA	GCC	GAC	ATC	GCC	2469
				775	Leu				780					785		
AGC	AAG	GTC	GCC	CAC	TCA	GCA	CTC	GTG	GAA	ACA	AGC	GAC	GCC	CTT	GAA	2517
Ser	Lys	Val	Ala 790	His	Ser	Ala	Leu	Val 795	Glu	Thr	Ser	Asp	800	Leu	Glu	
GCA	GTT	CAG	TCG	ACT	TCC	GTG	TAC	ACC	CCC	AAG	TAC	CCA	GAA	GŢC	AAG	2565
		805			Ser		810					815				
AAC	CCA	CAG	ACC	GCC	TCC	AAC	CCC	GTT	GTT	GGG	CIC	CAC	CTG	CCC.	GCC	2613
Asn	Pro 820	Gln	Thr	Ala	Ser	Asn 825	Pro	Val	Val	Gly	Leu 830	His	Leu	Pro	Ala	
AAG	AGA	GCC	ACC	GGT	GTC	CAG	GCC	GCT	CTT	CTC	GGA	GCA	GGA	ACG	AGC	2661
Lys	Arg	Ala	Thr	Gly	Val	Gln	Ala	Ala	Leu	Leu	Gly	Ala	Gly	Thr	Ser	. •
835					840					845					850	
AGA	CCA	ATG	GGG	ATG	GAG	GCC	CCA	ACA	CGG	TCC	AAG	AAC	GCC	GTG	AAA	2709
Arg	Pro	Met	Gly	Met 855	Glu	Ala	Pro	Tnr	860	ser.	гу	ASII	· HTG	865	руѕ	
ATG	GCC	AAA	CGG	CGG	CAA	CGC	CAA	AAG	GAG	AGC	CGC	TAA	CAGC	CAT		2755
Met	Ala	Lys	Arg 870	Arg	Gln	Arg	Gln	Lys 875	Glu	Ser	Arg			-		
GAT	GGA	ACC I	ACTC	AAGA	AG A	GGAC	ACTA	A TC	CCAG	ACCC	CGT	ATCC	CCG	GCCT'	rccct	2815
GCG	GGGG	ccc (CC	•					٠	•						2827

(2) INFORMATION FOR SEQ ID NO:26:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 878 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Met Ser Asp Ile Phe Asn Ser Pro Gln Ala Arg Ser Thr Ile Ser Ala 1 5 10 15

Ala Phe Gly Ile Lys Pro Thr Ala Gly Gln Asp Val Glu Glu Leu Leu

Ile Pro Lys Val Trp Val Pro Pro Glu Asp Pro Leu Ala Ser Pro Ser Arg Leu Ala Lys Phe Leu Arg Glu Asn Gly Tyr Lys Val Leu Gln Pro Arg Ser Leu Pro Glu Asn Glu Glu Tyr Glu Thr Asp Gln Ile Leu Pro Asp Leu Ala Trp Met Arg Gln Ile Glu Gly Ala Val Leu Lys Pro Thr Leu Ser Leu Pro Ile Gly Asp Gln Glu Tyr Phe Pro Lys Tyr Tyr Pro Thr His Arg Pro Ser Lys Glu Lys Pro Asn Ala Tyr Pro Pro Asp Ile Ala Leu Leu Lys Gln Met Ile Tyr Leu Phe Leu Gln Val Pro Glu Ala Asn Glu Gly Leu Lys Asp Glu Val Thr Leu Leu Thr Gln Asn Ile Arg Asp Lys Ala Tyr Gly Ser Gly Thr Tyr Met Gly Gln Ala Asn Arg Leu Val Ala Met Lys Glu Val Ala Thr Gly Arg Asn Pro Asn Lys Asp Pro Leu Lys Leu Gly Tyr Thr Phe Glu Ser Ile Ala Gln Leu Leu Asp Ile Thr Leu Pro Val Gly Pro Pro Gly Glu Asp Asp Lys Pro Trp Val Pro Leu Thr Arg Val Pro Ser Arg Met Leu Val Leu Thr Gly Asp Val Asp Gly Asp Phe Glu Val Glu Asp Tyr Leu Pro Lys Ile Asn Leu Lys Ser Ser Ser Gly Leu Pro Tyr Val Gly Arg Thr Lys Gly Glu Thr Ile Gly Glu Met Ile Ala Ile Ser Asn Gln Phe Leu Arg Glu Leu Ser Thr Leu

Leu Lys Gln Gly Ala Gly Thr Lys Gly Ser Asn Lys Lys Leu Leu

	290					295					300				
Ser 305	Met	Leu	Ser	Asp	Tyr 310	Trp	Tyr	Leu	Ser	Cys 315	Gly	Leu	Leu	Phe	Pro 320
Lys	Ala	Ģlu	Arg	Tyr 325	Asp	Lys	Ser	Thr	Trp 330	Leu	Thr	Lys	Thr	Arg 335	Asn
Ile	Trp	Ser	Ala 340	Pro	Ser	Pro	Thr	His 345	Leu	Met	Ile	Ser	Met 350	Ile	Thr
Trp	Pro	Val 355	Met	Ser	Asn	Ser	Pro 360	Asn	Asn	Val	Leu	Asn 365	Ile	Glu	Gly
Cys	Pro 370	Ser	Leu	Tyr	Lys	Phe 375	Asn	Pro	Phe	Arg	Gly 380	Gly	Leu	Asn	Arg
Ile 385	Val	Glu	Trp	Ile	Leu 390	Ala	Pro	Glu	Glu	Pro 395	Lys	Ala	Leu	Val	Tyr 400
Ala	Asp	Asn	Ile	Tyr 405	Ile	Val	His	Ser	Asn 410	Thr	Trp	Tyr	Ser	Ile 415	Asp
Leu	Glu	Lys	Gly 420	Glu	Ala	Asn	Cys	Thr 425	Arg	Gln	His	Met	Gln 430	Ala	Ala
	Tyr	435					440					445			
Phe	Asn 450	Gln	Thr	Trp	Ala	Thr 455	Phe _.	Ala	Met	Asn	Ile 460	Ala	Pro	Ala	Leu
Val 465	Val	Asp	Ser		Cys 470							Ile	Lys	Thr	Tyr 480
Gly	Gln	Gly	Ser	Gly 485	Asn	Ala	Ala	Thr	Phe 490	Ile	Asn	Asn	His	Leu 495	Leu
	Thr		500					505					510		
Asp	Ser	Glu 515	Glu	Phe	Lys	Ser	Ile 520	Glu	Asp	Lys	Leu	Gly 525	Ile	Asn	Phe
Lys	Ile 530	Glu	Arg	Ser	Ile	Asp 535	Asp	Ile	Arg	Gly	Lys 540	Leu	Arg	Gln	Leu
Val 545	Leu	Leu	Ala	Gln	Pro 550	Gly	Tyr	Leu	Ser	Gly 555	Gly	Val	Glu	Pro	Glu 560
Gln	Ser	Ser	Pro	Thr	Val	Glu	Leu	Asp	Leu	Leu	Gly	Trp	Ser	Ala	Thr

Tyr Ser Lys Asp Leu Gly Ile Tyr Val Pro Val Leu Asp Lys Glu Arq Leu Phe Cys Ser Ala Ala Tyr Pro Lys Gly Val Glu Asn Lys Ser Leu Lys Ser Lys Val Gly Ile Glu Gln Ala Tyr Lys Val Val Arg Tyr Glu Ala Leu Arg Leu Val Gly Gly Trp Asn Tyr Pro Leu Leu Asn Lys Ala Cys Lys Asn Asn Ala Gly Ala Ala Arg Arg His Leu Glu Ala Lys Gly Phe Pro Leu Asp Glu Phe Leu Ala Glu Trp Ser Glu Leu Ser Glu Phe Gly Glu Ala Phe Glu Gly Phe Asn Ile Lys Leu Thr Val Thr Ser Glu Ser Leu Ala Glu Leu Asn Lys Pro Val Pro Pro Lys Pro Pro Asn Val Asn Arg Pro Val Asn Thr Gly Gly Leu Lys Ala Val Ser Asn Ala Leu Lys Thr Gly Arg Tyr Arg Asn Glu Ala Gly Leu Ser Gly Leu Val Leu Leu Ala Thr Ala Arg Ser Arg Leu Gln Asp Ala Val Lys Ala Lys Ala Glu Ala Glu Lys Leu His Lys Ser Lys Pro Asp Asp Pro Asp Ala Asp Trp Phe Glu Arg Ser Glu Thr Leu Ser Asp Leu Leu Glu Lys Ala Asp 780 . . Ile Ala Ser Lys Val Ala His Ser Ala Leu Val Glu Thr Ser Asp Ala Leu Glu Ala Val Gln Ser Thr Ser Val Tyr Thr Pro Lys Tyr Pro Glu Val Lys Asn Pro Gln Thr Ala Ser Asn Pro Val Val Gly Leu His Leu

Pro Ala Lys Arg Ala Thr Gly Val Gln Ala Ala Leu Leu Gly Ala Gly

835 840 845

Thr Ser Arg Pro Met Gly Met Glu Ala Pro Thr Arg Ser Lys Asn Ala 850 855 860

Val Lys Met Ala Lys Arg Arg Gln Arg Gln Lys Glu Ser Arg 865 870 875

- (2) INFORMATION FOR SEQ ID NO:27:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3261 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: circular
 - (ii) MOLECULE TYPE: cDNA
 - (ix) FEATURE:

950

- (A) NAME/KEY: CDS
 - (B) LOCATION: 97..531
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

955

GGAT	ACGA	TC G	GTCI	GACC	C CC	GGGG	AGTO	ACC	CCGGG	GAC	AGGC	CCGTC	AA C	GCCI	TGTTC	60
CAGO	SATGO		CTCCI	CCTI	C TA	CAAC	CGCTA	A TC	ATTG			AGT Ser				114
ACA Thr 885	AAC Asn	GAT Asp	CGC Arg	AGC Ser	GAT Asp 890	GAC Asp	AAA Lys	CCT Pro	GCA Ala	AGA Arg 895	TCA Ser	AAC Asn	CCA Pro	ACA Thr	GAT Asp 900	162
TGT Cys	TCC Ser	GTT Val	CAT His	ACG Thr 905	GAG Glu	CCT Pro	TCT Ser	GAT Asp	GCC Ala 910	AAC Asn	AAC Asn	CGG Arg	ACC Thr	GGC Gly 915	Val	210
CAT His	TCC Ser	GGA Gly	CGA Arg 920	CAC His	CCT Pro	GGA Gly	GAA Glu	GCA Ala 925	CAC His	TCT Ser	CAG Gln	GTC Val	AGA Arg 930	GAC Asp	CTC Leu	258
GAC Asp	CTA Leu	CAA Gln 935	TTT Phe	GAC Asp	TGT Cys	GGG Gly	GGA Gly 940	CAC His	AGG Arg	GTC Val	AGG Arg	GCT Ala 945	AAT Asn	TGT Cys	CTT Leu	306
TTT Phe	CCC Pro	TGG Trp	ATT Ile	CCC Pro	TGG Trp	CTC Leu	AAT Asn	TGT Cys	GGG Gly	TGC Cys	TCA Ser	CTA Leu	CAC His	ACT Thr	GCA Ala	354

GGG CAA TGG GAA CTA CAA GTT CGA TCA GAT GCT CCT GAC TGC CCA GAA	402
Gly Gln Trp Glu Leu Gln Val Arg Ser Asp Ala Pro Asp Cys Pro Glu 965 970 975 980	
CCT ACC GGC CAG TTA CAA CTA CTG CAG GCT AGT GAG TCG GAG TCT CAC	450
Pro Thr Gly Gln Leu Gln Leu Gln Ala Ser Glu Ser Glu Ser His 985 990 995	••
AGT GAG GTC AAG CAC ACT TCC TGG TGG CGT TTA TGC ACT AAA CGG CAC	498
Ser Glu Val Lys His Thr Ser Trp Trp Arg Leu Cys Thr Lys Arg His 1000 1005 1010	
CAT AAA CGC CGT GAC CTT CCA AGG AAG CCT GAG TGAACTGACA GATGTTAGCT	551
His Lys Arg Arg Asp Leu Pro Arg Lys Pro Glu 1015 1020	
ACAATGGGTT GATGTCTGCA ACAGCCAACA TCAACGACAA AATTGGGAAC GTCCTAGTAG	611
GGGAAGGGT CACCGTCCTC AGCTTACCCA CATCATATGA TCTTGGGTAT GTGAGGCTTG	671
GTGACCCCAT TCCCGCAATA GGGCTTGACC CAAAAATGGT AGCCACATGT GACAGCAGTG	731
ACAGGCCCAG AGTCTACACC ATAACTGCAG CCGATGATTA CCAATTCTCA TCACAGTACC	791
AACCAGGTGG GGTAACAATC ACACTGTTCT CAGCCAACAT TGATGCCATC ACAAGCCTCA	851
GCGTTGGGGG AGAGCTCGTG TTTCAAACAA GCGTCCACGG CCTTGTACTG GGCGCCACCA	911
TCTACCTCAT AGGCTTTGAT GGGACAACGG TAATCACCAG GGCTGTGGCC GCAAACAATG	971
GGCTGACGAC CGGCACCGAC AACCTTATGC CATTCAATCT TGTGATTCCA ACAAACGAGA	1031
TAACCCAGCC AATCACATCC ATCAAACTGG AGATAGTGAC CTCCAAAAGT GGTGGTCAGG	1091
CAGGGGATCA GATGTCATGG TCGGCAAGAG GGAGCCTAGC AGTGACGATC CATGGTGGCA	1151
ACTATCCAGG GGCCCTCCGT CCCGTCACGC TAGTGGCCTA CGAAAGAGTG GCAACAGGAT	1211
CCGTCGTTAC GGTCGCTGGG GTGAGCAACT TCGAGCTGAT CCCAAATCCT GAACTAGCAA	1271
AGAACCTGGT TACAGAATAC GGCCGATTTG ACCCAGGAGC CATGAACTAC ACAAAATTGA	1331
TACTGAGTGA GAGGGACCGT CTTGGCATCA AGACCGTCTG GCCAACAAGG GAGTACACTG	1391
ACTTTCGTGA ATACTTCATG GAGGTGGCCG ACCTCAACTC TCCCCTGAAG ATTGCAGGAG	1451
CATTCGGCTT CAAAGACATA ATCCGGGCCA TAAGGAGGAT AGCTGTGCCG GTGGTCTCCA	1511
CATTGTTCCC ACCTGCCGCT CCCCTAGCCC ATGCAATTGG GGAAGGTGTA GACTACCTGC	1571
TGGGCGATGA GGCACAGGCT GCTTCAGGAA CTGCTCGAGC CGCGTCAGGA AAAGCAAGAG	1621

CTGCCTCAGG	CCGCATAAGG	CAGCTGACTC	TCGCCGCCGA	CAAGGGGTAC	GAGGTAGTCG	1691
CGAATCTATT	CCAGGTGCCC	CAGAATCCCG	TAGTCGACGG	GATTCTTGCT	TCACCTGGGG	1751
TACTCCGCGG	TGCACACAAC	CTCGACTGCG	TGTTAAGAGA	GGGTGCCACG	CTATTCCCTG	1811
TGGTTATTAC	GACAGTGGAA	GACGCCATGA	CACCCAAAGC	ATTGAACAGC	AAAATGTTTG	1871
CTGTCATTGA	AGGCGTGCGA	GAAGACCTCC	AACCTCCATC	TCAAAGAGGA	TCCTTCATAC	1931
GAACTCTCTC	TGGACACAGA	GTCTATGGAT	ATGCTCCAGA	TGGGGTACTT	CCACTGGAGA	1991
CTGGGAGAGA	CTACACCGTT	GTCCCAATAG	ATGATGTCTG	GGACGACAGC	ATTATGCTGT	2051
CCAAAGATCC	CATACCTCCT	ATTGTGGGAA	ACAGTGGAAA	TCTAGCCATA	GCTTACATGG	2111
ATGTGTTTCG	ACCCAAAGTC	CCAATCCATG	TGGCTATGAC	GGGAGCCCTC	AATGCTTGTG	2171
GCGAGATTGA	GAAAGTAAGC	TTTAGAAGCA	CCAAGCTCGC	CACTGCACAC	CGACTTGGCC	2231
TTAGGTTGGC.	TGGTCCCGGA	GCATTCGATG	TAAACACCGG	GCCCAACTGG	GCAACGTTCA	2291
TCAAACGTTT	CCCTCACAAT	CCACGCGACT	GGGACAGGCT	CCCCTACCTC	AACCTACCAT	2351
ACCTTCCACC	CAATGCAGGA	CGCCAGTACC	ACCTTGCCAT	GGCTGCATCA	GAGTTCAAAG	2411
AGACCCCCGA	ACTCGAGAGT	GCCGTCAGAG	CAATGGAAGC	AGCAGCCAAC	GTGGACCCAC	2471
TATTCCAATC	TGCACTCAGT	GTGTTCATGT	GGCTGGAAGA	GAATGGGATT	GTGACTGACA	2531
TGGCCAACTT	CGCACTCAGC	GACCCGAACG	CCCATCGGAT	GCGAAATTTT	CTTGCAAACG	2591
CACCACAAGC	AGGCAGCAAG	TCGCAAAGGG	CCAAGTACGG	GACAGCAGGC	TACGGAGTGG	2651
AGGCTCGGGG	CCCCACACCA	GAGGAAGCAC	AGAGGGAAAA	AGACACACGG	ATCTCAAAGA	2711
AGATGGAGAC	CATGGGCATC	TACTTTGCAA	CACCAGAATG	GGTAGCACTC	AATGGGCACC	2771
GAGGGCCAAG	CCCCGGCCAG	CTAAAGTACT	GGCAGAACAC	ACGAGAAATA	CCGGACCCAA	2831
ACGAGGACTA	TCTAGACTAC	GTGCATGCAG	AGAAGAGCCG	GTTGGCATCA	GAAGAACAAA	2891
TCCTAAGGGC	AGCTACGTCG	ATCTACGGGG	CTCCAGGACA	GGCAGAGCCA	CCCCAAGCTT	2951
TCATAGACGA	AGTTGCCAAA	GTCTATGAAA	TCAACCATGG	ACGTGGCCCA	AACCAAGAAC	3011
AGATGAAAGA	TCTGCTCTTG	ACTGCGATGG	AGATGAAGCA	TCGCAATCCC	AGGCGGGCTC	3071
TACCAAAGCC	CAAGCCAAAA	CCCAATGCTC	CAACACAGAG	ACCCCCTGGT	CGGCTGGGCC	3131
GCTGGATCAG	GACCGTCTCT	GATGAGGACC	TTGAGTGAGG	CTCCTGGGAG	TCTCCCGACA	3191

CCACCCGCGC AGGTGTGGAC ACCAATTCGG CCTTACAACA TCCCAAATTG GATCCGTTCG 3251
CGGGTCCCCT 3261

- (2) INFORMATION FOR SEQ ID NO:28:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 145 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Met Val Ser Arg Asp Gln Thr Asn Asp Arg Ser Asp Asp Lys Pro Ala 1 5 10 15

Arg Ser Asn Pro Thr Asp Cys Ser Val His Thr Glu Pro Ser Asp Ala 20 25 30

Asn Asn Arg Thr Gly Val His Ser Gly Arg His Pro Gly Glu Ala His 35 40 45

Ser Gln Val Arg Asp Leu Asp Leu Gln Phe Asp Cys Gly Gly His Arg 50 55 60

Val Arg Ala Asn Cys Leu Phe Pro Trp Ile Pro Trp Leu Asn Cys Gly
65 70 75 80

Cys Ser Leu His Thr Ala Gly Gln Trp Glu Leu Gln Val Arg Ser Asp 85 90 95

Ala Pro Asp Cys Pro Glu Pro Thr Gly Gln Leu Gln Leu Gln Ala 100 105 110

Ser Glu Ser Glu Ser His Ser Glu Val Lys His Thr Ser Trp Trp Arg

Leu Cys Thr Lys Arg His His Lys Arg Arg Asp Leu Pro Arg Lys Pro 130 135 140

Glu 145

- (2) INFORMATION FOR SEQ ID NO:29:
 - (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3261 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: circular
- (ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 131..3166

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

	,,,,,							_								
GGAT	ACGA	ATC C	GTCI	(GAC	CC CC	GGGG	SAGT	CAC	CCGG	GAC	AGG	CCGT	CAA (GCC:	TGTTC	60
CAGG	ATG	GA (TCC	CCT	C T	ACAAC	CGCT	A TC	ATTG/	ATGG	TTA	GTAG	AGA :	rcag,	ACAAAC	120
GATC	GCAG					eu (3ln (CAG 1 Gln 1 155				169
TTC :							Pro									217
GAC (Asp /																265
AAT '															Pro	313
GGA Gly																361
GGG .																409
GCC Ala																457
TCA Ser																505
GCC	GTG	ACC	TTC	CAA	GGA	AGC	CTG	AGT	GAA	CTG	ACA	GAT	GTT	AGC	TAC	553

Ala	Val	Thr	Phe	Gln 275	Gly	Ser	Leu	Ser	Glu 280	Leu	Thr	Asp	Val	Ser 285	Tyr	
														GGG Gly		601
														TCA Ser		649
														GGG Gly		697
														AGA Arg		745
														TAC Tyr 365		793
									Ser					GCC Ala		841
														GTC Val		889
														GGG Gly		937
														ACC Thr	_	985
														GAG Glu 445		1033
														AAA Lys		1081
														AGC Ser		1129

			ATC Ile													1177
ACG Thr 495	CTA Leu	GTG Val	GCC Ala	TAC Tyr	GAA Glu 500	AGA Arg	GTG Val	GCA Ala	ACA Thr	GGA Gly 505	TCC Ser	GTC Val	GTT Val	ACG Thr	GTC Val 510	1225
GCT Ala	GGG Gly	GTG Val	AGC Ser	AAC Asn 515	TTC Phe	GAG Glu	CTG Leu	ATC Ile	CCA Pro 520	AAT Asn	CCT Pro	GAA Glu	CTA Leu	GCA Ala 525	AAG Lys	1273
AAC Asn	CTG Leu	GTT Val	ACA Thr 530	GAA Glu	TAC Tyr	GGC Gly	CGA Arg	TTT Phe 535	GAC Asp	CCA Pro	GGA Gly	GCC Ala	ATG Met 540	AAC Asn	TAC Tyr	1321
ACA Thr	AAA Lys	TTG Leu 545	ATA Ile	CTG Leu	AGT	GAG Glu	AGG Arg 550	GAC Asp	CGT Arg	CTT	GGC Gly	ATC Ile 555	AAG Lys	ACC Thr	GTC Val	1369
TGG Trp	CCA Pro 560	ACA Thr	AGG Arg	GAG Glu	Tyr	ACT Thr 565	GAC Asp	TTT Phe	CGT Arg	GAA Glu	TAC Tyr 570	TTC Phe	ATG Met	GAG Glu	GTG Val	1417
GCC Ala 575	GAC Asp	CTC Leu	AAC Asn	TCT Ser	CCC Pro 580	CTG Leu	AAG Lys	ATT Ile	GCA Ala	GGA Gly 585	GCA Ala	TTC Phe	GGC Gly	TTC Phe	AAA Lys 590	1465
GAC Asp	ATA Ile	ATC Ile	CGG Arg	GCC Ala 595	ATA Ile	AGG Arg	AGG Arg	Ile	GCT Ala 600	GTG Val	CCG Pro	GTG Val	Val	TCC Ser 605	ACA Thr	1513
TTG Leu	TTC	CCA Pro	CCT Pro 610	Ala	GCT Ala	CCC Pro	CTA Leu	GCC Ala 615	CAT His	GCA Ala	ATT Ile	GGG Gly	GAA Glu 620	GGT Gly	GTA Val	1561
GAC Asp	TAC Tyr	CTG Leu 625	CTG Leu	GGC Gly	GAT Asp	GAG Glu	GCA Ala 630	CAG Gln	GCT Ala	GCT Ala	TCA Ser	GGA Gly 635	ACT Thr	GCT Ala	CGA Arg	1609
GCC Ala	GCG Ala 640	TCA Ser	GGA Gly	AAA Lys	GCA Ala	AGA Arg 645	GCT Ala	GCC Ala	TCA Ser	GGC Gly	CGC Arg 650	ATA Ile	AGG Arg	CAG Gln	CTG Leu	1657
ACT Thr 655	Leu	GCC Ala	GCC Ala	GAC Asp	AAG Lys 660	GGG	TAC Tyr	GAG Glu	GTA Val	GTC Val 665	GCG Ala	AAT Asn	CTA Leu	TTC Phe	CAG Gln 670	1705
GTG Val	CCC	CAG Gln	AAT Asn	CCC Pro 675	Val	GTC Val	GAC A sp	GGG	ATT Ile 680	Leu	GCT Ala	TCA Ser	CCT Pro	GGG Gly 685	GTA Val	1753

CTC Leu	CGC	GGT	GCA Ala 690	CAC	AAC Asn	CTC Leu	GAC Asp	TGC Cys 695	GTG Val	TTA Leu	AGA Arg	GAG Glu	GGT Gly 700	GCC Ala	ACG Thr	1801
Leu	Phe	Pro 705	Val	Val	Ile	Thr	Thr 710	Val	Glu	Asp	Ala	Met 715	Thr	Pro	AAA Lys	1849
Ala	Leu 720	Asn	Ser	Lys		Phe 725	Ala	Val	Ile	Glu	Gly 730	Val	Arg	Glu	Asp	1897
CTC Leu 735	CAA Gln	CCT	CCA Pro	TCT Ser	CAA Gln 740	AGA Arg	GGA Gly	TCC Ser	TTC Phe	ATA Ile 745	CGA Arg	ACT Thr	CTC Leu	TCT	GGA Gly 750	1945
CAC His	AGA Arg	GTC Val	TAT Tyr	GGA Gly 755	TAT	GCT Ala	CCA Pro	gat Asp	GGG Gly 760	GTA Val	CTT Leu	CCA Pro	CTG Leu	GAG Glu 765	ACT Thr	1993
		Asp			GTT Val											2041
					GAT Asp								Asri-			2089
					TAC Tyr											2137
					GGA Gly 820											2185
					ACC Thr											2233
					GGA Gly											2281
					CGT Arg	Phe					Arg					2329
					CTA Leu		Tyr									2377

TAC Tyr 895	CAC His	CTT Leu	GCC Ala	ATG Met	GCT Ala 900	GCA Ala	TCA Ser	GAG Glu	TTC Phe	AAA Lys 905	GAG Glu	ACC Thr	CCC Pro	GAA Glu	CTC Leu 910	2425 .
Glu	Ser	Ala	Val	AGA Arg 915	Ala	Met	Glu	Ala	Ala 920	Ala	Asn	Val	Asp	Pro 925	Leu	2473
Phe	Gln	Ser	Ala 930		Ser	Val	Phe	Met 935	Trp	Leu	Glu	Glu	Asn 940	Gly	Ile	2521
Val	Thr	Asp 945	Met	GCC Ala	Asn	Phe	Ala 950	Leu	Ser	Asp	Pro	Asn 955	Ala	His	Arg	2569
ATG Met	CGA Arg 960	AAT Asn	TTT Phe	CTT Leu	GCA Ala	AAC Asn 965	GCA Ala	CCA Pro	CAA Gln	GCA Ala	GGC Gly 970	AGC Ser	AAG Lys	TCG Ser	CAA Gln	2617
AGG Arg 975	GCC Ala	AAG Lys	TAC Tyr	GGG Gly	ACA Thr 980	GCA Ala	GGC Gly	TAC Tyr	GGA Gly	GTG Val 985	GAG Glu	GCT Ala	CGG Arg	GGC Gly	CCC Pro 990	2665
ACA Thr	CCA Pro	GAG Glu	GAA Glu	GCA Ala 995	CAG Gln	AGG Arg	GAA Glu	AAA Lys	GAC Asp 100	Thr	CGG Arg	ATC Ile	TCA Ser	AAG Lys 100	Lys	.2713
ATG Met	GAG Glu	ACC	ATG Met 101	GGC Gly O	ATC Ile	TAC Tyr	TTT Phe	GCA Ala 101	Thr	CCA Pro	GAA Glu	TGG	GTA Val 102	Ala	CTC Leu	2761
AAT Asn	GGG Gly	CAC His 102	Arg	GGG Gly	CCA Pro	AGC Ser	CCC Pro 103	Gly	CAG Gln	CTA Leu	AAG Lys	TAC Tyr 103	Trp	CAG Gln	AAC Asn	2809
ACA Thr	CGA Arg 104	Glu	ATA	CCG Pro	GAC Asp	Pro	Asn	GAG Glu	GAC Asp	TAT	CTA Leu 105	Asp	TAC	GTG Val	CAT His	2857
GCÁ Ala 105	Glu	AAG Lys	AGC Ser	CGG Arg	TTG Leu 106	Ala	TCA Ser	GAA Glu	GAA Glu	CAA Gln 106	Ile	CTA Leu	AGG Arg	GCA Ala	GCT Ala 1070	2905
ACG Thr	TCG Ser	ATC Ile	TAC	GGG Gly 107	Ala	CCA Pro	GGA Gly	CAG Gln	GCA Ala 108	Glu	CCA Pro	CCC	CAA Gln	GCT Ala 108	Phe	2953
ATA Ile	GAC Asp	GAA Glu	GTT Val 109	Ala	AAA Lys	GTC Val	TAT Tyr	GAA Glu 109	Ile	AAC	CAT His	GGA Gly	CGT Arg 110	Gly	CCA Pro	3001

								4	16							
AAC Asn	CAA Gln	GAA Glu 1105	Gln	ATG Met	AAA Lys	GAT Asp	CTG Leu 1110	Leu	TTG Leu	ACT Thr	GCG Ala	ATG Met	Glu	ATG Met	AAG Lys	3049
CAT His	CGC Arg 1120	Asn	CCC Pro	AGG Arg	CGG Arg	GCT Ala 1125	CTA Leu	CCA Pro	AAG Lys	CCC Pro	AAG Lys 1130	Pro	AAA Lys	CCC Pro	AAT Asn	3097
GCT Ala 1135	Pro	ACA Thr	CAG Gln	AGA Arg	CCC Pro 1140	Pro	GGT Gly	CGG Arg	CTG Leu	GGC Gly 1145	Arg	TGG Trp	ATC Ile	AGG Arg	ACC Thr 1150	3145
GTC Val	TCT Ser	GAT Asp	GAG Glu	GAC Asp 1155	Leu	GAG Glu	TGAG	GCTC	CT 6	GGAG	TCTC	CC CC	ACAC	CACC	2	3196
CGCG	CAGO	etg 1	GGAC	CACCA	A TI	CGGC	CTTA	CAA	CATO	CCA	AATI	'GGAT	'CC G	TTCG	CGGGT	3256
CCCC	T											•				3261
(2)			EQUE	NCE	CHAR	ACTE	0:30	ICS:					,	٠.		
			(B)	TYP	E: a		2 am aci		aC1Q	.S				• . •		

- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Met Thr Asn Leu Gln Asp Gln Thr Gln Gln Ile Val Pro Phe Ile Arg

Ser Leu Leu Met Pro Thr Thr Gly Pro Ala Ser Ile Pro Asp Asp Thr 30

Leu Glu Lys His Thr Leu Arg Ser Glu Thr Ser Thr Tyr Asn Leu Thr

Val Gly Asp Thr Gly Ser Gly Leu Ile Val Phe Pro Gly Phe Pro 50 55

Gly Ser Ile Val Gly Ala His Tyr Thr Leu Gln Gly Asn Gly Asn Tyr 65 70

Lys Phe Asp Gln Met Leu Leu Thr Ala Gln Asn Leu Pro Ala Ser Tyr 85 90 95

Asn Tyr Cys Arg Leu Val Ser Arg Ser Leu Thr Val Arg Ser Ser Thr 100 105 110

- Leu Pro Gly Gly Val Tyr Ala Leu Asn Gly Thr Ile Asn Ala Val Thr 115 120 125
- Phe Gln Gly Ser Leu Ser Glu Leu Thr Asp Val Ser Tyr Asn Gly Leu 130 135 140
- Met Ser Ala Thr Ala Asn Ile Asn Asp Lys Ile Gly Asn Val Leu Val 145 150 155 160
- Gly Glu Gly Val Thr Val Leu Ser Leu Pro Thr Ser Tyr Asp Leu Gly 165 170 175
- Tyr Val Arg Leu Gly Asp Pro Ile Pro Ala Ile Gly Leu Asp Pro Lys 180 185 190
- Met Val Ala Thr Cys Asp Ser Ser Asp Arg Pro Arg Val Tyr Thr Ile 195 200 205
- Thr Ala Ala Asp Asp Tyr Gln Phe Ser Ser Gln Tyr Gln Pro Gly Gly 210 215 220
- Val Thr Ile Thr Leu Phe Ser Ala Asn Ile Asp Ala Ile Thr Ser Leu 225 230 235 240
- Ser Val Gly Glu Leu Val Phe Gln Thr Ser Val His Gly Leu Val 245 250 255
- Leu Gly Ala Thr Ile Tyr Leu Ile Gly Phe Asp Gly Thr Thr Val Ile
 260 265 270
- Thr Arg Ala Val Ala Ala Asn Asn Gly Leu Thr Thr Gly Thr Asp Asn 275 280 285
- Leu Met Pro Phe Asn Leu Val Ile Pro Thr Asn Glu Ile Thr Gln Pro 290 295 300
- Ile Thr Ser Ile Lys Leu Glu Ile Val Thr Ser Lys Ser Gly Gln 305 310 315 320
- Ala Gly Asp Gln Met Ser Trp Ser Ala Arg Gly Ser Leu Ala Val Thr 325 330 335
- Ile His Gly Gly Asn Tyr Pro Gly Ala Leu Arg Pro Val Thr Leu Val 340 345 350
- Ala Tyr Glu Arg Val Ala Thr Gly Ser Val Val Thr Val Ala Gly Val 355 360 365
- Ser Asn Phe Glu Leu Ile Pro Asn Pro Glu Leu Ala Lys Asn Leu Val 370 375 380
- Thr Glu Tyr Gly Arg Phe Asp Pro Gly Ala Met Asn Tyr Thr Lys Leu

385	ì				390					395					400
Ile	Leu	Ser	Glu	Arg 405	Asp	Arg	Leu	Gly	Ile 410		Thr	Val	Trp	Pro 415	Thr
Arg	Glu	Tyr	Thr 420	Asp	Phe	Arg	Glu	Tyr 425		Met	Glu	Val	Ala 430	_	Leu
Asn	Ser	Pro 435	Leu	Lys	Ile	Ala	Gly 440	Ala	Phe	Gly	Phe	Lys		Ile	Ile
Arg	Ala 450	Ile	Arg	Arg	Ile	Ala 455	Val	Pro	Val	Val	Ser 460	Thr	Leu	Phe	Pro
Pro 465	Ala	Ala	Pro	Leu	Ala 470	His	Ala	Ile	Gly	Glu 475	Gly	Val	Asp	Tyr	Leu 480
Leu	Gly	Asp	Glu	Ala 485	Gln	Ala	Ala	Ser	Gly 490	Thr	Ala	Arg	Ala	Ala 495	Ser
Gly	Lys	Ala	Arg 500	Ala	Ala	Ser	Gly	Arg 505	Ile	Arg	Gln	Leu	Thr 510	Leu	Ala
Ala	Asp	Lys 515	Gly	Tyr	Glu	Val	Val 520	Ala	Asn	Leu	Phe	Gln 525	Val	Pro	Gln
Asn	Pro 530	Val	Val	Asp	Gly	Ile 535	Leu	Ala	Ser	Pro	Gly 540	Val	Leu	Arg	Gly
Ala 545	His	Asn	Leu	Asp	Cys 550	Val	Leu	Arg	Glu	Gly 555	Ala	Thr	Leu		Pro 560
Val	Val	Ile	Thr	Thr 565	Val	Glu	Asp	Ala	Met 570	Thr	Pro	Lys	Ala	Leu 575	Asn
Ser	Lys	Met	Phe 580	Ala	Val	Ile	Glu	Gly 585	Val	Arg	Glu	Asp	Leu 590	Gln	Pro
Pro	Ser	Gln 595	Arg	Gly	Ser	Phe	Ile 600	Arg	Thr	Leu	Ser	Gly 605	His	Arg	Val
Tyr	Gly 610	Tyr	Ala	Pro	Asp	Gly 615	Val	Leu	Pro		Glu .620	Thr	Gly	Arg	Asp
Tyr 625	Thr	Val	Val	Pro	Ile 630	Asp	Asp	Val	Trp	Asp 635	Asp	Ser	Ile		Leu 640
Ser	Lys	Asp	Pro	Ile 645	Pro	Pro	Ile	Val	Gly 650	Asn	Ser	Gly	Asn	Leu 655	Ala
Ile	Ala	Tyr	Met 660	Asp	Val	Phe	Arg	Pro 665	Lys	Val	Pro	Ile	His 670	Val	Ala

- Met Thr Gly Ala Leu Asn Ala Cys Gly Glu Ile Glu Lys Val Ser Phe 675 680 685
- Arg Ser Thr Lys Leu Ala Thr Ala His Arg Leu Gly Leu Arg Leu Ala 690 695 700
- Gly Pro Gly Ala Phe Asp Val Asn Thr Gly Pro Asn Trp Ala Thr Phe 705 710 715 720
- Ile Lys Arg Phe Pro His Asn Pro Arg Asp Trp Asp Arg Leu Pro Tyr 725 730 735
- Leu Asn Leu Pro Tyr Leu Pro Pro Asn Ala Gly Arg Gln Tyr His Leu 740 745 750
- Ala Met Ala Ala Ser Glu Phe Lys Glu Thr Pro Glu Leu Glu Ser Ala 755 760 765
- Val Arg Ala Met Glu Ala Ala Ala Asn Val Asp Pro Leu Phe Gln Ser 770 775 780
- Ala Leu Ser Val Phe Met Trp Leu Glu Glu Asn Gly Ile Val Thr Asp 785 790 795 800
- Met Ala Asn Phe Ala Leu Ser Asp Pro Asn Ala His Arg Met Arg Asn 805 810 815
- Phe Leu Ala Asn Ala Pro Gln Ala Gly Ser Lys Ser Gln Arg Ala Lys 820 825 830
- Tyr Gly Thr Ala Gly Tyr Gly Val Glu Ala Arg Gly Pro Thr Pro Glu 835 840 845
- Glu Ala Gln Arg Glu Lys Asp Thr Arg Ile Ser Lys Lys Met Glu Thr 850 855 860
- Met Gly Ile Tyr Phe Ala Thr Pro Glu Trp Val Ala Leu Asn Gly His 865 870 875 880
- Arg Gly Pro Ser Pro Gly Gln Leu Lys Tyr Trp Gln Asn Thr Arg Glu 885 890 895
- Ile Pro Asp Pro Asn Glu Asp Tyr Leu Asp Tyr Val His Ala Glu Lys 900 905 910
- Ser Arg Leu Ala Ser Glu Glu Gln Ile Leu Arg Ala Ala Thr Ser Ile 915 920 925
- Tyr Gly Ala Pro Gly Gln Ala Glu Pro Pro Gln Ala Phe Ile Asp Glu 930 935 940
- Val Ala Lys Val Tyr Glu Ile Asn His Gly Arg Gly Pro Asn Gln Glu

945				•	950					955					960	•
Gln	Met	Lys	Asp	Leu 965	Leu	Leu	Thr	Ala	Met 970	Glu	Met	Lys	His	Arg 975	Asn	
Pro	Arg	Arg	Ala 980	Leu	Pro	Lys	Pro	Lys 985	Pro	Lys	Pro	Asn	Ala 990	Pro	Thr	
Gln	Arg	Pro 995	Pro	Gly	Arg	Leu	Gly 1000	Arg	Trp	Ile	Arg	Thr 100		Ser	Asp	
Glu	Asp 1010		Glu			-										•
(2)	INFO	RMAT	NOI	FOR	SEQ	ID N	10:31	l:								
	(i)				IARAC										·	•
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		(0	:) S7	TRANI	DEDNE	ESS:	sing	jle								
		(E)) TC	POLC	GY:	circ	ular									,
	(ii)	MOI	ECUI	E T	PE:	CDNA	1									
•	(ix)		TURE											•		
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	(XI)	SEQ	OENC		ESCRI	PTIC	N: S	PEÖ 1	טא ע.):31:						
GGAI	'ACGA	TC G	GTCI	GACC	CC CG	GGGG	AGTO	ACC	CGGG	GAC	AGG	CATO	AC I	GCCI	TGTTC	60
CTGG	TTGG	AA C	TCCI	CTTI	C TG	CTGT	ACTA	TCG	TTG	ATG	GTG	AGT	AGA	GAT	CAG	114
												Ser	Arg			
												1015				•• .
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					GAT Asp											162
	1020		3			1025			- I F	- 2	1030		110	****	wah	
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					Glu											210
1035					1040					1045	•			-	1050	
CAT	TCC	GGA	CGA	CAC	CCT	GGA	GAA	GCA	CAC	ACT	CAG	GTC	CGA	AAC	CTC	258
			Arg	His	Pro				His	Thr				Asn	Leu	
				1055	•				1060)				1065		
GAC	TTA	CAA	CTT	GAC	TGT	AGG	GGA	TAC	AGG	GTC	AGG	ACT	AAT	TGT	CTT	306

Asp	Leu	Gln	Leu 1070		Cys	Arg	Gly	Tyr 1075		Val	Arg	Thr	Asn 108		Leu		
TTT Phe	CCC Pro	TGG Trp 1085	Ile	CCC Pro	TGG Trp	TTC Phe	AGT Ser 1090	Сув	AGG Arg	TGC Cys	TCA Ser	CTA Leu 1099	His	ACT Thr	GCA Ala	,	354
Glu	CAG Gln 1100	Trp	GAA Glu	CTA Leu	CCA Pro	ATT Ile 1105	Arg	CCA Pro	GAT Asp	GCT Ala	CCT Pro 1110	Asp	AGC Ser	GCA Ala	GAA Glu		402
CCT Pro 1115	Ala	TGC Cys	CAG Gln	CTA Leu	CAA Gln 1120	Leu	CTG Leu	CAG Gln	GCT Ala	AGT Ser 1125	Glu	CAG Gln	GAG Glu	TCT	AAC Asn 1130	•	450
CGT Arg	ACG Thr	GTC Val	AAG Lys	CAC His	Thr	CCC Pro	TGG Trp	TGG Trp	CGT Arg 1140	Leu	TGC Cys	ACT Thr	AAA Lys	CGG Arg 114	Asn	٠.	498
			AGT Ser 1150	Asp					Pro		TGA	GTTG/	ACT	GACT	ACAGC	r	551
ACAA	CGGG	CT (ATG	CAGO	CC AC	CTGCC	BAAC	TC	AACG/	ACAA	GAT	CGGG	AAC	GTTC	TAGTT	3	611
GAGA	AGG	GT (ACTO	TTC	rc Ac	TCT?	ACCG	A CT	CAT	ATGA	CCT	ragt:	TAT	GTGA	GACTC	3	671
GTGA	CCCC	CAT (cccc	CAGO	CA GO	SACTO	CGAC	C CGI	AAGT	rgat	GGC	CACG'	rgc	GACA	GTAGT	3	731
ACAG	ACC	CAG A	AGTCT	racac	CC AT	raac:	AGCTO	G CAC	GATG	AATA	CCA	ATTC"	rcg	TCAC	AACTC	A	791
TCCC	GAGI	rgg (CGTG	AAGA	CC AC	CACTO	STTC1	r cco	GCCA!	ACAT	CGA'	rgct	CTC	ACCA	GCTTC	A	851
GCGI	TGG	rgg :	rgago	CTTG:	rc T	rcago	CCAA	G TA	ACGA!	rcca	AAG	CATT	GAA	GTGG.	ACGTC	A	911
CCAT	TCA	CTT (CATT	GGT.	rr G	ACGG	GACA	G AC	GTAG	CAGT	CAA	GCA	GTT	GCAA	CAGAC	T	971
TTG	GCT	GAC 2	AACTO	GGA(CA A	ACAA	CCTT	G TG	CCAT	TCAA	CCT	GGTG	GTC	CCAA	CAAAT	G :	1031
AGAT	CAC	CCA (GCCC2	ATCA	CT TO	CAT	GAAA	C TAC	GAGG'	PTGT	GAC	CTAC	AAG	ATTG	GCGGC.	A :	1091
CCG	TGG	rga (CCCA	ATAT(CA TO	GGAC	AGTG	A GT	GGTA	CACT	AGC"	rgtg.	ACG	GTGC	ACGGA	G :	1151
GCA	ACTA	CCC '	TGGG	GCTC'	rc c	GTCC'	TGTC	A CC	CTGG'	rggc	CTA'	TGAA	CGA	GTGG	CTGCA	G·:	1211
GATO	TGT	rgt (CACA	GTTG	CA G	GGGT	GAGC	a ac'	TTCG	AGCT	AAT	CCCC.	AAC	CCTG	AGCTT	G :	1271
CAA	AGAA(CCT /	AGTT	ACAG	AG T	ATGG	CCGC	T TT	GACC	CCGG	AGC.	AATG.	AAC	TÀCA	CCAAA	C	133:
TAA	ract(GAG '	TGAG	AGAG.	AT C	GTCT.	AGGC	A TC.	AAGA	CAGT	CTG	GCCC	ACC	AGGG	AGTAC	A :	1391
CCG	ATTT	CAG ·	GGAG'	TACT'	TC A	TGGA	GGTT	G CA	GATC	TCAA	CTC	ACCC	CTA	AAGA	TTGCA	G	1451

GAGCATITGG	CITIAAGGAC	ATAATCCGAG	CCATTCGGAA	GATTGCGGTG	CCAGTGGTAT	1511
CCACACTCTT	CCCTCCAGCT	GCACCCCTAG	CACATGCAAT	CGGAGAAGGT	GTAGACTACC	1571
TCCTGGGCGA	CGAGGCCCAA	GCAGCCTCAG	GGACAGCTCG	AGCCGCGTCA	GGAAAAGCTA	1631
GAGCTGCCTC	AGGACGAATA	AGGCAGCTAA	CTCTCGCAGC	TGACAAGGGG	TGCGAGGTAG	1691
TCGCCAACAT	GTTCCAGGTG	CCCCAGAATC	CCATTGTTGA	TGGCATTCTG	GCATCCCCAG	1751
GAATCCTGCG	TGGCGCACAC	AACCTCGACT	GCGTGCTATG	GGAGGGAGCC	ACTCTTTTCC	1811
CTGTTGTCAT	TACGACACTC	GAGGATGAGC	TGACCCCCAA	GGCACTGAAC	AGCAAAATGT	1871
TTGCTGTCAT	TGAAGGTGTG	CGAGAGGACC	TCCAGCCTCC	ATCCCAACGG	GGATCCTTCA	1931
TTCGAACTCT	CTCTGGCCAT	AGAGTCTATG	GCTATGCCCC	AGACGGAGTA	CTGCCTCTGG	1991
AGACCGGGAG	AGACTACACC	GTTGTCCCAA	TTGATGATGT	GTGGGACGAT	AGCATAATGC	2051
TGTCGCAGGA	CCCCATACCT	CCAATCATAG	GGAACAGCGG	CAACCTAGCC	ATAGCATACA	2111
TGGATGTCTT	CAGGCCCAAG	GTCCCCATCC	ACGTGGCTAT	GACAGGGGCC	CTCAATGCCC	2171
GCGGTGAGAT	CGAGAGTGTT	ACGTTCCGCA	GCACCAAACT	CGCCACAGCC	CACCGACTTG	2231
GCATGAAGTT	AGCTGGTCCT	GGAGCCTATG	ACATTAATAC	AGGACCTAAC	TGGGCAACGT	2291
rcgtcaaacg	TTTCCCTCAC	AATCCCCGAG	ACTGGGACAG	GTTGCCCTAC	CTCAACCTTC	2351
CTTATCTCCC	ACCAACAGCA	GGACGTCAGT	TCCATCTAGC	CCTGGCTGCC	TCCGAGTTCA	2411
AAGAGACCCC	AGAACTCGAA	GACGCTGTGC	GCGCAATGGA	TGCCGCTGCA	AATGCCGACC	2471
CATTGTTCCG	CTCAGCTCTC	CAGGTCTTCA	TGTGGTTGGA	AGAAAACGGG	ATTGTGACCG	2531
ACATGGCTAA	CTTCGCCCTC	AGCGACCCAA	ACGCGCATAG	GATGAAAAAC	TTCCTAGCAA	2591
ACGCACCCCA	GGCTGGAAGC	AAGTCGCAGA	GGGCCAAGTA	TGGCACGGCA	GGCTACGGAG	2651
rggaggctcg	AGGCCCCACA	CCAGAAGAGG	CACAGAGGGA	AAAAGACACA	CGGATCTCCA	2711
AGAAGATGGA	AACAATGGGC	ATCTACTTCG	CGACACCGGA	ATGGGTGGCT	CTCAACGGGC	2771
ACCGAGGCCC	AAGCCCCGGC	CAACTCAAGT	ACTGGCAAAA	CACAAGAGAA	ATACCAGAGC	2831
CCAATGAGGA	CTACCCAGAC	TATGTGCACG	CGGAGAAGAG	CCGGTTGGCG	TCAGAAGAAC	2891
AGATCCTACG	GGCAGCCACG	TCGATCTACG	GGGCTCCAGG	ACAGGCTGAA	CCACCCCAGG	2951
CCTTCATAGA	CGAGGTCGCC	AGGGTCTATG	AAATCAACCA	TGGGCGTGGT	CCAAACCAGG	3011

AGCAGATGAA	GGACCTGCTC	CTGACTGCGA	TGGAGATGAA	GCATCGCAAT	CCCAGGCGGG	3071
CTCCACCAAA	GCCAAAGCCA	AAACCCAATG	CTCCATCACA	GAGACCCCCT	GGACGCTGG .	3131
GCCGCTGGAT	CAGGACGGTC	TCCGACGAGG	ACTTGGAGTG	AGGCTCCTGG	GAGTCTCCCG	3191
ACACTACCCG	CGCAGGTGTG	GACACCAATT	CGGCCTTCTA	CCATCCCAAA	TTGGATCCGT	3251
TCGCGGGTCC	CCT			•		3264

(2) INFORMATION FOR SEQ ID NO:32:

WU 98/09040

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 145 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:
- Met Val Ser Arg Asp Gln Thr Asn Asp Arg Ser Asp Asp Lys Pro Asp
 1 5 10 15
- Gly Ser His Pro Thr Asp Cys Ser Val His Thr Glu Pro Ser Asp Ala 20 25 30
- Asn Asp Arg Thr Gly Val His Ser Gly Arg His Pro Gly Glu Ala His
 35 40 45
- Thr Gln Val Arg Asn Leu Asp Leu Gln Leu Asp Cys Arg Gly Tyr Arg 50 55 60
- Val Arg Thr Asn Cys Leu Phe Pro Trp Ile Pro Trp Phe Ser Cys Arg
 65 70 75 80
- Cys Ser Leu His Thr Ala Glu Gln Trp Glu Leu Pro Ile Arg Pro Asp 85 90 95
- Ala Pro Asp Ser Ala Glu Pro Ala Cys Gln Leu Gln Leu Gln Ala
 100 105 110
- Ser Glu Gln Glu Ser Asn Arg Thr Val Lys His Thr Pro Trp Trp Arg
- Leu Cys Thr Lys Arg Asn His Lys Arg Ser Asp Leu Pro Arg Lys Pro
 130 135 140

Glu

(2) INFORMATION FOR SEQ ID NO:33:

 	CHARACTERISTICS:

(A) LENGTH: 3264 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: circular

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 131..3169

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

			•	
GGATACGATC	GGTCTGACCC CGGG	GGAGTC ACCCGGGGAC	AGGCCATCAC TGCCTTGTTC	60
CTGGTTGGAA	CTCCTCTTTC TGCT	STACTA TCGTTGATGG	TGAGTAGAGA TCAGACAAAC	120
GATCGCAGCG			CAA CAG ATT GTT CCG Gln Gln Ile Val Pro 155	169
		t Pro Thr Thr Gly	CCG GCG TCC ATT CCG Pro Ala Ser Ile Pro 170	217
			GAA ACC TCG ACT TAC Glu Thr Ser Thr Tyr 190	265
			ATT GTC TTT TTC CCT Ile Val Phe Phe Pro 205	313
			ACA CTG CAG AGC AGT Thr Leu Gln Ser Ser 220	361
	Gln Phe Asp Gl		GCG CAG AAC CTG CCT Ala Gln Asn Leu Pro 235	409
		g Leu Val Ser Arg	AGT CTA ACC GTA CGG Ser Leu Thr Val Arg 250	457

			CTC Leu								Asn						505
			TTC Phe														553
			ATG Met 290														601
			GGA Gly														649
			TAT Tyr												CTC Leu	٠.	697
			TTG Leu														745
TAC Tyr	ACC Thr	ATA Ile	ACA Thr	GCT Ala 355	GCA Ala	GAT Asp	GAA Glu	TAC	CAA Gln 360	TTC Phe	TCG Ser	TCA Ser	CAA Gln	CTC Leu 365	Ile		793
CCG Pro	AGT Ser	GGC	GTG Val 370	AAG Lys	ACC Thr	ACA Thr	CTG Leu	TTC Phe 375	TCC Ser	GCC Ala	AAC Asn	ATC Ile	GAT Asp 380	GCT Ala	CTC		841
ACC	Ser	TTC Phe 385	AGC Ser	GTT Val	GGT Gly	GGT Gly	GAG Glu 390	CTT Leu	GTC Val	TTC Phe	AGC Ser	CAA Gln 395	GTA Val	ACG Thr	ATC Ile		889
			GAA Glu														937
ACA Thr 415	GAC Asp	GTA Val	GCA Ala	GTC Val	AAG Lys 420	GCA Ala	GTT Val	GCA Ala	ACA Thr	GAC Asp 425	TTT Phe	GGG Gly	CTG Leu	ACA Thr	ACT Thr 430		985
			AAC Asn													:	1033
			CCC Pro 450														1081

				GAC Asp					ACA Thr	1129
				GGA Gly 485						1177
				GAA Glu						1225
				TTC Phe						1273
				TAT Tyr					AAC Asn	1321
				AGT Ser						1369
				TAC Tyr 565						1417
				CCC Pro						1465
		Ile		ATT Ile						1513
				GCA Ala						1561
				GAC Asp						1609
				GCT Ala 645						1657
	_			AAG Lys						1705

CAG Gln	GTG Val	CCC Pro	CAG Gln	AAT Asn 675	CCC Pro	ATT Ile	GTT Val	GAT Asp	GGC Gly 680	ATT Ile	CTG Leu	GCA Ala	TCC Ser	CCA Pro 685	GGA Gly	1753
ATC Ile	CTG Leu	CGT Arg	GGC Gly 690	GCA Ala	CAC His	AAC Asn	CTC Leu	GAC Asp 695	TGC Cys	GTG Val	CTA Leu	TGG Trp	GAG Glu 700	GGA Gly	GCC Ala	1801
ACT	CTT Leu	TTC Phe 705	CCT Pro	GTT Val	GTC Val	ATT Ile	ACG Thr 710	ACA Thr	CTC Leu	GAG Glu	GAT Asp	GAG Glu 715	Leu	ACC Thr	CCC	1849
AAG Lys	GCA Ala 720	CTG Leu	AAC Asn	AGC Ser	AAA Lys	ATG Met 725	TTT Phe	GCT Ala	GTC Val	ATT Ile	GAA Glu 730	GGT Gly	GTG Val	CGA Arg	GAG Glu	1897
GAC Asp 735	CTC Leu	CAG Gln	CCT Pro	CCA Pro	TCC Ser 740	CAA Gln	CGG Arg	GGA Gly	TCC Ser	TTC Phe 745	ATT Ile	CGA Arg	ACT	CTC Leu	TCT Ser 750	1945
GGC	CAT His	AGA Arg	GTC Val	TAT Tyr 755	GC	TAT Tyr	GCC Ala	CCA Pro	GAC Asp 760	GGA Gly	GTA Val	CTG Leu	CCT Pro	CTG Leu 765	GAG Glu	1993
ACC Thr	GGG Gly	AGA Arg	GAC Asp 770	TAC Tyr	ACC Thr	GTT Val	Val	CCA Pro 775	ATT Ile	GAT Asp	GAT Asp	GTG Val	TGG Trp 780	GAC Asp	GAT Asp	2041
AGC Ser	ATA Ile	ATG Met 785	CTG Leu	TCG Ser	CAG Gln	GAC Asp	CCC Pro 790	ATA Ile	CCT Pro	CCA Pro	ATC Ile	ATA Ile 795	GGG Gly	AAC Asn	AGC Ser	2089
GGC	AAC Asn 800	CTA Leu	GCC Ala	ATA	GCA Ala	TAC Tyr 805	ATG Met	GAT Asp	GTC Val	TTC	AGG Arg 810	CCC Pro	AAG Lys	GTC Val	Pro	2137
ATC Ile 815	CAC His	GTG Val	GCT Ala	ATG Met	ACA Thr 820	GGG Gly	GCC	CTC Leu	AAT Asn	GCC Ala 825	CGC	GGT	GAG Glu	ATC Ile	GAG Glu 830	2185
AGT Ser	GTT Val	ACG Thr	TTC Phe	CGC Arg 835	AGC Ser	ACC Thr	AAA Lys	CTC	GCC Ala 840	ACA Thr	GCC Ala	CAC His	CGA Arg	CTT Leu 845	GGC Gly	2233
ATG Met	AAG Lys	TTA Leu	GCT Ala 850	GGT Gly	CCT	GGA Gly	GCC Ala	TAT Tyr 855	GAC Asp	ATT	AAT Asn	ACA Thr	GGA Gly 860	CCT	AAC Asn	2281
TGG Trp	GCA Ala	ACG Thr 865	TTC	GTC Val	AAA Lys	CGT	TTC Phe 870	CCT	CAC	AAT Asn	CCC	CGA Arg 875	GAC Asp	TGG Trp	GAC Asp	2329

AGG Arg	TTG Leu 880	CCC	TAC	CTC Leu	AAC Asn	CTT Leu 885	CCT Pro	TAT	CTC	CCA Pro	CCA Pro 890	Thr	GCA Ala	GGA Gly	CGT Arg	2377
Gln 895	Phe	His	Leu	Ala	Leu 900	Ala	Ala	Ser	Glu	Phe 905	Lys	Glu	Thr	Pro	910	2425
CTC Leu	GAA Glu	GAC	GCT Ala	GTG Val 915	CGC Arg	GCA Ala	ATG Met	GAT Asp	GCC Ala 920	GCT	GCA Ala	AAT	GCC Ala	GAC Asp 925	CCA Pro	2473
TTG Leu	TTC Phe	CGC Arg	TCA Ser 930	GCT Ala	CTC Leu	CAG Gln	GTC Val	TTC Phe 935	ATG Met	TGG Trp	TTG Leu	GAA Glu	GAA Glu 940	AAC Asn	GGG Gly	2521
ATT	GTG Val	ACC Thr 945	GAC Asp	ATG Met	GCT Ala	AAC Asn	TTC Phe 950	GCC Ala	CTC Leu	AGC Ser	GAC Asp	CCA Pro 955	AAC Asn	GCG Ala	CAT His	2569
AGG Arg	ATG Met 960	AAA Lys	AAC Asn	TTC Phe	CTA Leu	GCA Ala 965	AAC Asn	GCA Ala	CCC	CAG Gln	GCT Ala 970	GGA Gly	AGC Ser	AAG Lys	TCG Ser	2617
			AAG Lys													2665
			GAA Glu							qaA					Lys	2713
AAG Lys	ATG Met	GAA Glu	ACA Thr 1010	Met	GGC Gly	Ile	TAC	TTC Phe 1015	Ala	ACA Thr	CCG Pro	GAA Glu	TGG Trp 1020	Val	GCT Ala	2761
CTC Leu	AAC Asn	GGG Gly 1025		CGA Arg	GGC Gly	CCA Pro	AGC Ser 1030	Pro	GGC Gly	CAA Gln	CTC Leu	AAG Lys 1035	Tyr	TGG Trp	CAA Gln	2809
		Arg	GAA Glu				Pro					Pro				2857
	Ala		AAG Lys			Leu			Glu		Gln			Arg		2905
			ATC Ile		Gly					Ala					Ala	2953

TTC Phe	ATA Ile	GAC Asp	GAG Glu 1090	Val	GCC Ala	AGG Arg	GTC Val	TAT Tyr 1095	Glu	ATC Ile	AAC Asn	CAT His	GGG Gly 1100	Arg	GGT Gly	3001
CCA Pro	AAC Asn	CAG Gln 1105	Glu	CAG Gln	ATG Met	AAG Lys	GAC Asp 1110	Leu	CTC Leu	CTG Leu	ACT Thr	GCG Ala 1115	Met	GAG Glu	ATG Met	3049
AAG Lys	CAT His 1120	Arg	AAT Asn	CCC Pro	AGG Arg	CGG Arg 1125	Ala	CCA Pro	CCA Pro	AAG Lys	CCA Pro 1130	Lys	CCA Pro	AAA Lys	CCC Pro	3097
AAT Asn 113	Ala	CCA Pro	TCA Ser	CAG Gln	AGA Arg 1140	Pro	CCT Pro	GGA Gly	CGG Arg	CTG Leu 1145	Gly	CGC Arg	TGG Trp	ATC Ile	AGG Arg 1150	3145
			GAC Asp		Asp			TGAG	GCT(CT C	GGAG	FTCTC	C CG	SACA(CTACC	3199
CGC	GCA G(TG :	rggao	CACC	TT AA	rcgg(CCTT	TAC	CATO	CCCA	AATT	rgga?	rcc c	TTC	SCGGGT	3259
CCC	CT.	•													•	3264
(2)	INF	ORMA!	rion	FOR	SEQ	ID I	NO:34	<u>.</u>	•					. •		• •• •
		(i)	(A)	LEI	NGTH:	: 10: amin	ERIST 13 am o act line	nino id	e acio	ls						
	(ii)	MOLE	CULE	TYPI	E: p:	rote	in			. •				. •	
	(:	xi)	SEQU	ENCE	DES	CRIP'	TION	: SE	Q ID	NO:	3 4 :		٠.	,	•	
Met 1	Thr	Asn	Leu	Met 5	Asp	His	Thr	Gln	Gln 10	Ile	Val	Pro	Phe	Ile 15	Arg	
Ser	Leu	Leu	Met 20	Pro	Thr	Thr	Gly	Pro 25	Ala	Ser	Ile	Pro	Asp 30	Asp	Thr	
Leu	Glu	Lys 35		Thr	Leu	Arg	Ser 40	Glu	Thr	Ser	Thr	Tyr 45	Asn	Leu	Thr	
Val	Gly 50		Thr	Gly	Ser	Gly 55		Ile	Val	Phe	Phe 60	Pro	Gly	Phe	Pro	
Gly 65		Val	Val	Gly	Ala 70		Tyr	Thr	Leu	Gln 75	Ser	Ser	Gly	Asn	Tyr 80	
Glr	Phe	Asp	Gln	Met	Leu	Leu	Thr	Ala	Gln	Asn	Leu	Pro	Ala	Ser	Tyr	

Asn	Tyr	Сув	Arg 100	Leu	Val	Ser	Arg	Ser 105	Leu	Thr	Val	Arg	Ser 110		Thr
Leu	Pro	Gly 115	Gly	Val	Tyr	Ala	Leu 120	Asn	Gly	Thr	Ile	Asn 125	Ala	Val	Thr
Phe	His 130	Gly	Ser	Leu	Ser	Glu 135	Leu	Thr	Asp	Tyr	Ser 140	Tyr	Asn	Gly	Leu
Met 145	Ser	Ala	Thr	Ala	Asn 150	Ile	Asn	Asp	Lys	Ile 155	Gly	Asn	Val	Leu	Val 160
Gly	Glu	Gly	Val	Thr 165	Val	Leu	Ser	Leu	Pro 170	Thr	Ser	Tyr	Asp	Leu 175	Ser
Tyr	Val	Arg	Leu 180	Gly	Asp	Pro	Ile	Pro 185	Ala	Ala	Gly	Leu	Asp 190	Pro	Lys
Leu	Met	Ala 195	Thr	Cys	Asp	Ser	Ser 200	Asp	Arg	Pro	Arg	Val 205	Tyr	Thr	Ile
Thr	Ala 210	Ala	qaA	Glu	Tyr	Gln 215	Phe	Ser	Ser	Gln	Leu 220	Ile	Pro	Ser	Gly
Val 225	Lys	Thr	Thr	Leu	Phe 230	Ser	Ala	Asn	Ile	Asp 235	Ala	Leu	Thr	Ser	Phe 240
Ser	Val	Gly	Gly	Glu 245	Leu	Val	Phe	Ser	Gln 250	Val	Thr	Ile	Gln	Ser 255	Ile
Glu	Val	Asp	Val 260	Thr	Ile	His	Phe	Ile 265	Gly	Phe	Asp	Gly	Thr 270	Asp	Val
Ala	Val	Lys 275	Ala	Val	Ala	Thr	Asp 280	Phe	Gly	Leu	Thr	Thr 285	Gly -	Thr	Asn
	Leu 290	Val	Pro	Phe	Asn	Leu 295	Val	Val	Pro	Thr	Asn 300	Glu	Ile	Thr	Gln
Pro 305	Ile	Thr	Ser	Met	Lys 310	Leu	Glu	Val		Thr 315	Tyr	Lys	Ile	Gly	Gly 320
Thr	Ala	Gly	Asp	Pro 325	Ile	Ser	Trp .	Thr	Val 330	Ser	Gly	Thr	Leu	Ala 335	Val
fhr	Val		Gly 340	Gly	Asn	Tyr	Pro	Gly 345	Ala	Leu	Arg	Pro	Val 350	Thr	Leu
/al		Tyr 355	Glu	Arg	Val	Ala	Ala 360	Gly	Ser	Val	Val	Thr 365	Val	Ala	Gly

- Val Ser Asn Phe Glu Leu Ile Pro Asn Pro Glu Leu Ala Lys Asn Leu 370 380
- Val Thr Glu Tyr Gly Arg Phe Asp Pro Gly Ala Met Asn Tyr Thr Lys 385 390 395
- Leu Ile Leu Ser Glu Arg Asp Arg Leu Gly Ile Lys Thr Val Trp Pro 405 410 415
- Thr Arg Glu Tyr Thr Asp Phe Arg Glu Tyr Phe Met Glu Val Ala Asp 420 425 430
- Leu Asn Ser Pro Leu Lys Ile Ala Gly Ala Phe Gly Phe Lys Asp Ile 435 440 445
- Ile Arg Ala Ile Arg Lys Ile Ala Val Pro Val Val Ser Thr Leu Phe 450 455 460
- Pro Pro Ala Ala Pro Leu Ala His Ala Ile Gly Glu Gly Val Asp Tyr 465 470 475 480
- Leu Leu Gly Asp Glu Ala Gln Ala Ala Ser Gly Thr Ala Arg Ala Ala 485 490 495
- Ser Gly Lys Ala Arg Ala Ala Ser Gly Arg Ile Arg Gln Leu Thr Leu 500 505 510
- Ala Ala Asp Lys Gly Cys Glu Val Val Ala Asn Met Phe Gln Val Pro 515 520 525
- Gln Asn Pro Ile Val Asp Gly Ile Leu Ala Ser Pro Gly Ile Leu Arg
 530 535 540
- Gly Ala His Asn Leu Asp Cys Val Leu Trp Glu Gly Ala Thr Leu Phe 545 550 555 560
- Pro Val Val Ile Thr Thr Leu Glu Asp Glu Leu Thr Pro Lys Ala Leu 565 570 575
- Asn Ser Lys Met Phe Ala Val Ile Glu Gly Val Arg Glu Asp Leu Gln 580 585 590
- Pro Pro Ser Gln Arg Gly Ser Phe Ile Arg Thr Leu Ser Gly His Arg 595 600 605
- Val Tyr Gly Tyr Ala Pro Asp Gly Val Leu Pro Leu Glu Thr Gly Arg 610 620
- Asp Tyr Thr Val Val Pro Ile Asp Asp Val Trp Asp Asp Ser Ile Met 625 635 640

Leu	Ser	Gln	Asp	Pro 645	Ile	Pro	Pro	Ile	Ile 650	Gly	Asn	Ser	Gly	Asn 655	Leu
Ala	Ile	Ala	Tyr 660	Met	Asp	Val	Phe	Arg 665	Pro	Lys	Val	Pro	Ile 670	His	Val
Ala	Met	Thr 675	Gly	Ala	Leu	Asn	Ala 680	Arg	Gly	Glu	Ile	Glu 685	Ser	Val	Thr
Phe	Arg 690	Ser	Thr	Lys	Leu	Ala 695	Thr	Ala	His	Arg	Leu 700	Gly	Met	Lys	Leu
Ala 705		Pro	Gly	Ala	Tyr 710	Asp	Ile	Asn	Thr	Gly 715	Pro	Asn	Trp	Ala	Thr 720
Phe	Val	Lys	Arg	Phe 725	Pro	His	Asn	Pro	Arg 730	Asp	Trp	Asp	Arg	Leu 735	Pro
Tyr	Leu	Asn	Leu 740	Pro	Tyr	Leu	Pro	Pro 745	Thr	Ala	Gly	Arg	Gln 750	Phe	His
Leu	Ala	Leu 755	Ala	Ala	Ser	Glu	Phe 760	Lys	Glu	Thr	Pro	Glu 765	Leu	Glu	Asp
Ala	Val 770	Arg	Ala	Met	Asp	Ala 775	Ala	Ala	Asn	Ala	Asp 780	Pro	Leu	Phe	Arg
Ser 785	Ala	Leu	Gln	Val	Phe 790	Met	Trp	Leu	Glu	Glu 795	Asn	Gly	Ile	Val	Thr 800
Asp	Met	Ala	Asn	Phe 805	Ala	Leu	Ser	Asp	Pro 810	Asn	Ala	His	Arg	Met 815	Lys
Asn	Phe	Leu	Ala 820	Asn	Ala	Pro	Gln	Ala 825	Gly	Ser	Lys	Ser	Gln 830	Arg	Ala
Lys	Tyr	Gly 835	Thr	Ala	Gly	Tyr	Gly 840	Val	Glu	Ala	Arg	Gly 845	Pro	Thr	Pro
Glu	Glu 850	Ala	Gln	Arg	Glu	Lys 855	Asp	Thr	Arg	Ile	Ser 860	Lys	Lys	Met	Glu
Thr 865	Met	Gly	Ile	Tyr	Phe 870	Ala	Thr	Pro	Glu	Trp 875	Val	Ala	Leu	Asn	Gly 880
His	Arg	Gly	Pro	Ser 885	Pro	Gly	Gln	Leu	Lys 890	Tyr	Trp	Gln	Asn	Thr 895	Arg
Glu	Ile	Pro	Glu 900	Pro	Asn	Glu	Asp	Tyr 905	Pro	Asp	Tyr	Val	His 910	Ala	Glu

- Lys Ser Arg Leu Ala Ser Glu Glu Gln Ile Leu Arg Ala Ala Thr Ser 915 920 925
- Ile Tyr Gly Ala Pro Gly Gln Ala Glu Pro Pro Gln Ala Phe Ile Asp 930 935 940
- Glu Val Ala Arg Val Tyr Glu Ile Asn His Gly Arg Gly Pro Asn Gln 945 950 955 960
- Glu Gln Met Lys Asp Leu Leu Leu Thr Ala Met Glu Met Lys His Arg 965 970 975
- Asn Pro Arg Ala Pro Pro Lys Pro Lys Pro Lys Pro Asn Ala Pro 980 985 990
- Ser Gln Arg Pro Pro Gly Arg Leu Gly Arg Trp Ile Arg Thr Val Ser 995 1000 1005

Asp Glu Asp Leu Glu 1010

Claims

1. A method for preparing live Birnavirus, comprising the following steps:

preparing a cDNA containing infectious bursal disease virus genome segments A and B,

transcribing said cDNA to produce synthetic RNA transcripts, transfecting host cells with said synthetic RNA transcripts, incubating said host cells in a culture medium, and isolating live infectious bursal disease virus from said culture medium.

- 2. The method according to claim 1, wherein said Birnavirus is infectious bursal disease virus.
- 3. The method according to claim 1, wherein said host cells are African green monkey Vero cells.
- 4. The method according to claim 1, wherein said segments A and B of said cDNA are independently prepared.
- 5. The method according to claim 4, wherein said segment A is present in plasmid pUC19FLAD78 or pUC18FLA23.
- 6. The method according to claim 4, wherein said segment B is present in plasmid pUC18FLBP2.
- 7. A live infectious bursal disease virus, wherein said virus is made by a process comprising the steps of preparing a cDNA containing infectious bursal disease virus genome segments A and B,

transcribing said cDNA to produce a synthetic RNA transcript, transfecting a host cell with said synthetic RNA transcript, incubating said host cell in a culture medium, and isolating live infectious bursal disease virus from said culture medium.

- 8. A synthetic RNA encoding proteins VP1, VP2, VP3, VP4, and VP5 of infectious bursal disease virus.
 - 9. A host cell transfected with the synthetic RNA according to claim 8.
- 10. A cDNA containing at least a portion of the infectious bursal disease virus genome selected from the group consisting of segment A,

segment B and segments A and B of infectious bursal disease virus, wherein said cDNA includes the 5' and 3' terminii of said segments.

- 11. A recombinant vector comprising the cDNA according to claim 10.
- 12. The vector according to claim 11, wherein said vector is a plasmid.
- 13. The vector according to claim 12, wherein said plasmid is selected from the group consisting of pUC19FLAD78, pUC18FLA23 and pUC19FLBP2.
 - 14. A host cell transformed with the vector according to claim 11.
- 15. A vaccine comprising an infectious bursal disease virus according to claim 7, wherein said infectious bursal disease virus is inactivated or attenuated prior to administration.
- 16. A method for producing a live infectious bursal disease virus vaccine, comprising the steps of

preparing a full-length cDNA containing infectious bursal disease virus genome segments A and B,

transcribing said cDNA to produce synthetic RNA transcripts, purifying said synthetic RNA transcripts, transfecting host cells with said purified RNA transcripts,

incubating said host cells in a culture medium,

isolating live infectious bursal disease virus from said culture medium, attenuating said live infectious bursal disease virus to produce a virus with reduced virulence, and

combining said live infectious bursal disease virus with a pharmaceutically acceptable carrier to produce a live infectious bursal disease virus vaccine.

- 17. The method according to claim 16, wherein said live infectious bursal disease virus is attenuated by serial passage or site directed mutagenesis.
- 18. The method according to claim 1, wherein said host cells are poultry cells.
- 19. The method according to claim 18, wherein said poultry cells are chicken, turkey, or quail cells.

20. The method according to claim 19, wherein said poultry cells are chicken embryo fibroblast cells or chicken embryo kidney cells.

Fig. 1

Fig. 4

Fig. 5

Fig. 6

Fig. IA

Fig. IB

Fig. IC

Fig. 4A

Fig.4B

Fig. 5A

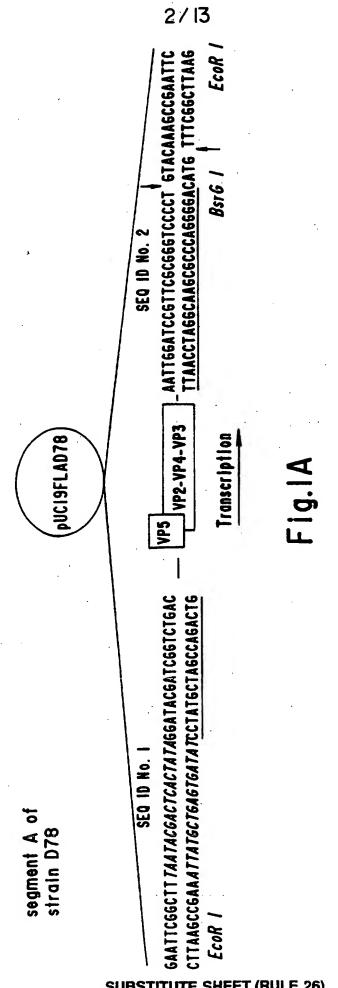
Fig. 5B

Fig. 6A

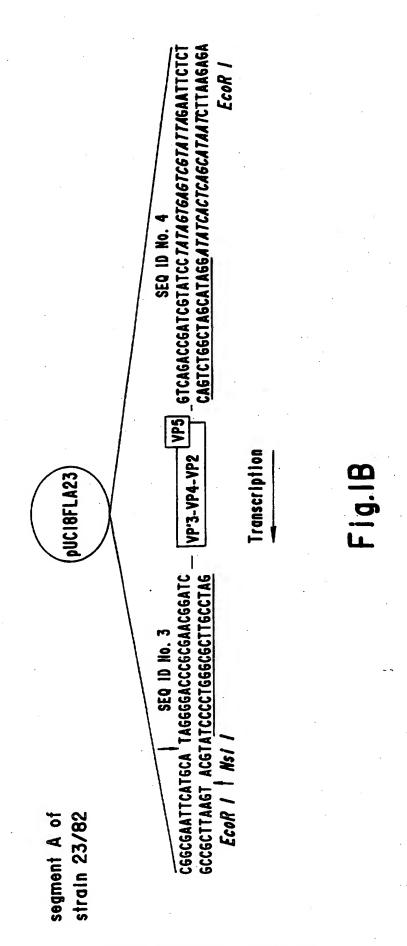
Fig. 6B

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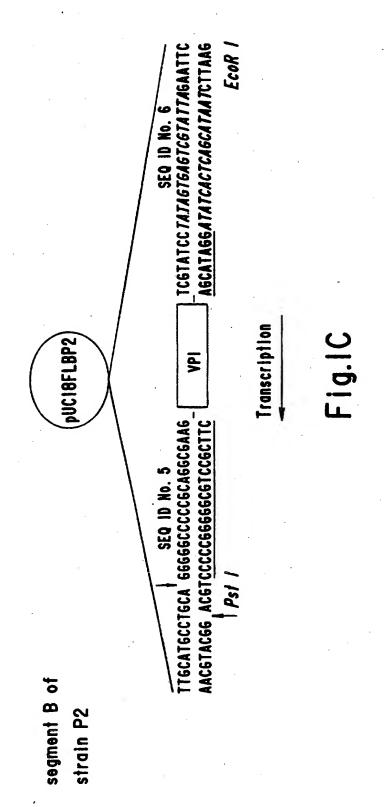
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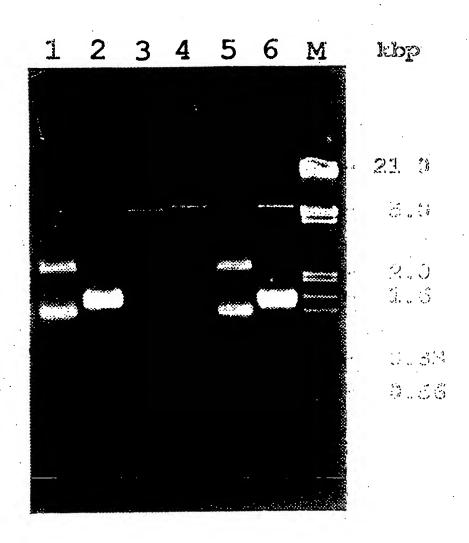


Fig. 2

V
3
D)
_

	530	540	550	260	570	280
23-82A	66AAGCCT6AG1	GGAAGCCTGAGTGAGTTGACTGACTACAGCTACAACGGGCTGATGTCAGCCACTGCGAAC	TACAGCTAC	ACGGCTGA	TETCAGCCACT	GCGAAC
23A/P28	GGAAGCCTGAGI	66AAGCCTGAGTGAGTTGACTGACTACAGCTACAACGGGCTGATGTCAGCCACTGCGAAC	TACAGCTAC	AACGGGCTGA	TETCAGCCACT	GCGAAC
SEQ ID NO. 8 P2A SEQ ID No. 9	66AAGCCTGAGT 530	66AAGCCTGAGTGACAGATGTTAGCTACAATGGGTTGATGTCTGCAACAGCCAAC 530 540 550 560 570 580	GTTAGCTAC	AATGGGTTGA 560	TAGCTACAATGGGTTGATGTCTGCAACAGCCAA 550 560 570 58	GCCAAC 580
23-82A SFO 10 No 7	590 ATCAACGACAA	590 630 610 610 620 630 640 ATCAACGACAAGATCGGGAACGTTCTAGTTGGAGAGGGGTGACTGTTCTAGTTGGAGAGGGGTGACTGTTCTAGTCTACCG	610 ICTAGTTGGA	620 6AGGGTGA	630 CTGTTCTCAG	640 CTACC6
23A/P2B SF0 10 No. 8	ATCAACGACAA	ATCAACGACAAGATCGGGAACGTTCTAGTTGGAGGGGTGACTGTTCTCAGTCTACCG	CTAGTTGGA	SAAGGGTGA	CTETTCTCAG	CTACCE
P2A SEQ ID No. 9	ATCAACGACAA/	ATCAACGACAAATTGGGAACGTCCTAGTAGGGGAAGGGGTCACCGTCCTCAGCTTACCC 590 600 610 640	CTAGTAGGG 610	6A6666TCA 620	CCETCCTCAGG 630	TTACCC 640

Segment A

egment B

23-82B	9	130 140 150 160 160 170 180 TTTTCAATAGECETAGECETAGECETAGE	140 CACAGGGGGA	150 ACGAGATC	160 TCAGCAGCGT	170 TCGGCATAAA	180 IECCTACTE
23A/P28		TTTCAACAGTCCACAGGCGCGAAGCACGATCTCAGCAGCGTTCGGCATAAAGCCTACTG	CACAGGCGCGA	AGCACGATC	CACGATCTCAGCAGCGTTCGGC/	rceecataaa	GCCTACT6
P28 SEQ 10 No. 12	- 2	TTTTCAACAGTCCACAGGGGGGGGTCTCAGCGGTTCGGCGTAAAGCCTACTG	CACAGGGGGAAG	AGCACGATC 150	ICAGCAGCGT1 160	CGGCATAAA 170	GCCTACTG 180
23-828 20 10 10 10	9	190 CTEGACAGACE	200 TEGAAGAACTC	210 TTGATCCCC	220 NAAGTCT6661	230 TGCCACCTGA	240 66ATCC6C
23A/P28	2	CT66ACA6ACGA6AACTCTT6ATCCCTAAAGTTT666T6CCACCT6A6GATCC6C	TEGAAGAACTC	TTGATCCCT	AAGTTT666	TECCACCTEA	66ATCC6C
SEQ ID No. 11 P2B	=	CTGGACAAGACGTGGAAGTCCCTAAAGTTTGGGTGCCACCTGAGGATCCGC	TEGAGGACTC	TTGATCCCT	AA6111666	Feccace Fea	66ATCC6C
SEQ 1D No. 12	2	190	190 200	210	220	230	240

Fig.3B

GGGAGTACTTCATGGAGGTTGCAGATCTCAACTCACCCCTAAAGATTGCAGGAGCATTTGGCTTTAAGGA ACCATTCACTTCATTGGGTTTGACGGGACAGACGTAGCAGTCAAGGCAGTTGCAACAGACTTTGGGCTGA TCCATGAAACTAGAGGTTGTGACCTACAAGATTGGCGGCACCGCTGGTGACCCAATATCATGGACAGTG A GTGGTA CACTAGCT GTGA CGG TGCA CGGA GCCAA CTAC CCTGGGGCT CTCCGT CCTGTCAC CCTGGTGG **CCCTGAGCTTGCAAAGAACCTAGTTACAGAGTATGGCCGCTTTGACCCCGGAGCAATGAACTACACCAAA** CTAATACTGAGTGAGAGATCGTCTAGGCATCAAGACAGTCTGGCCCACCAGGGAGTACACCGATTTGA CATAATCCGAGCCATTCGGAAGATTGCGGTGCCAGTGGTATCCACACTCTTCCCTCCAGCTGCACCCCTA **SCACATGCAATCGGAGAAGGTGTAGACTACCTCCTGGGCGACGAGGCCCAAGCAGCCTCAGGGACAGCTC** GAGCCGCGTCAGGAAAAGCTAGAGCTGCCTCAGGACGAATAAGGCAGCTAACTCTCGCAGCTGACAA GGG CAACTGGGACAAACAACCTTGTGCCATTCAACCTGGTGGTCCCAACAAATGAGATCACCCAGCCCATCAC **CCTATGAACGAGTGGCTGCAGGATCTGTTGTCACAGTTGCAGGGGTGAGCAACTTCGAGCTAATCCCCAA GTGCGAGGTAGTCGCCAACATGTTCCAGGTGCCCCAGAATCCCATTGTTGATGGCATTCTGGCATCCCCA** CATTCCGGACGACACCCTGGAGAGCACACACTCAGGTCCGAAACCTCGACTTACAACTTGACTGTAGGG **CCGAAGTTGATGGCCACGTGCGACAGTAGTGACAGACCCAGAGTCTACACCATAACAGCTGCAGATGAAT** CACCAGCTTCAGCGTTGGTGGTGAGCTTGTCTTCAGCCAAGTAACGATCCAAAGCATTGAAGTGGACGTC **CTACTGCAGGCTAGTGAGCAGGAGTCTAACCGTACGGTCAAGCACACTCCCTGGTGGCGTTTATGCACTA** TGATGTCAGCCACTGCGAACATCAACGACAAGATCGGGAACGTTCTAGTTGGAGAAGGGGTGACTGTTCT **CAGTCTACCGACTTCATATGACCTTAGTTATGTGAGACTCGGTGACCCCATCCCCGCAGCAGGACTCGAC GGATACGATCGGTCTGACCCCGGGGAGTCACCCGGGGACAGGCCATCACTGCCTTGTTCCTGGTTGGAA GATACAGGGTCAGGACTAATTGTCTTTTTCCCTGGATTCCCTGGTTCAGTTGTAGGTGCTCACTACACAC** CTCCTCTTTCTGCTGTACTATCGTTGATGGTGAGAGATCAGACAAACGATCGCAGCGATGACAAACC IGA TGGA T CACACCCAACAGA T TGT T C GT T CATACGGA G C CTT C T GA T G C CAA C G A C C G G C C G C G T C **ACCAATTCTCGTCACAACTCATCCCGAGTGGCGTGAAGACCACACTGTTCTCCGCCAACATGGTGCTCT** 8 771 841 40 47 98 261 331 5 55 49 561 63 <u>=</u>6 2 <u>6</u> <u>=</u> 35 42 281 4

TCCATCTAGCCCTGGCTGCCTCCGAGTTCAAAGAGACCCCAGAACTCGAAGACGCTGTGCGCGCAATGG ATGCCGCTGCAAATGCCGACCCATTGTTCCGCTCAGCTCTCCAGGTCTTCATGTGGTTGGAAGAAACGG **GATTGTGACCGACATGGCTAACTTCGCCCTCAGCGACCCAAACGCGCATAGGATGAAAAACTTCCTAGCA GAGGCCCCACACCAGAGAGGCACAGAGGGAAAAAGACACGGATCTCCAAGAAGATGGAAACAATGGG** CATCTACTTCGCGACACCGGAATGGGTGGCTCTCAACGGGCACCGAGGCCCAAGCCCCGGCCAACTCAAG **TACTGGCAAAACACAAGAGAATACCAGAGCCCAATGAGGACTACCCAGACTATGTGCACGCGGAGAAGA GCCGGTTGGCGTCAGAAGAACAGATCCTACGGGCAGCCACGTCGATCTACGGGGCTCCAGGACAGGCTGA** accacccc aggccttcatagacgaggtcgccagggtctatgaaatcaaccatgggcgtggtccaaacca **SAGCAGATGAAGGACCTGCTCCTGACTGCGATGGAGGATCAACCATCGCAATCCCAGGCGGGCTCCACCAA AGCCAAAGCCAAAACCCAATGCTCCATCACAGAGCCCCCTGGACGGCTGGGCCGCTGGATCAG GACGGT** CTCCGACG AGGACTTGGAGTGAGGCTCCTGGGAGTCTCCCGACACTACCCGCGCGGGTGT GGACACCAAT **AACGCACCCCAGGCTGGAAGCAAGTCGCAGAGGGCCAAGTATGGCACGGCAGGCTACGGAGTGGAGGCTC** 5GCTATGCCCCAGACGGAGTACTGCCTCTGGAGACCGGGAGAGTACACCGTTGTCCCAATTGATGATG **AGCTGGTCCTGGAGCCTATGACATTAATACAGGACCTAACTGGGCAACGTTCGTCAAACGTTTCCCTCA** |GTGGGACGATAGCATAATGCTGTCGCAGGACCCCATACCTCCAATCATAGGGAACAGCGGCAACCTAGC CATAGCATACATGGATGTCTTCAGGCCCAAGGTCCCCATCCACGTGGCTATGACAGGGGCCCTCAATGCC ;GCGGTGAGATCGAGAGTGTTACGTTCCGVAGCACCAAACTCGCCACAGCCCACCGACTTGGCATGAAGT SCGAGAGGACCTCCAGCCTCCATCCCAACGGGGATCCTTCATTCGAACTCTCTGGCCATAGAGTCTA TACGACACTCGAGGATGAGCTGACCCCCAAGGCACTGAACAGCAAAATGTTTGCTGTCATTGAAGGTG **ICGG CCTTCTACCATCCCAAATTGGATCCGTTCGCGGGTCCCCT** 2941 2451 2591 2661 2731 2801 301 3081 2381 2521 287 3151 2241 231 196 203 2101 2171

Total number of bases is: 3264. DNA sequence composition: 834 A; 942 C; 853 G;

635

Sequence name: 23-82A (SEQ ID NOS: 31 and 33

Fig.4B

CATGCAATTGGGGAAGGTGTAGACTACCTGCTGGGCGATGAGGCACAGGCTGCTTCAGGAACTGCTCGAG CGAGGTAGTCGCGAATCTATTCCAGGTGCCCCAGAATCCCGTAGTCGACGGGATTCTTGCTTCACCTGGG STACTCCGCGGTGCACACACCTCGACTGCGTGTTAAGAGGGGGGCCCCCCCTATTCCCTGTGGTTATTA TGCAGGGCAATGGGAACTACAAGTTCGATCAGATGCTCCTGACTGCCCAGAACCTACCGGCCAGTTACAA CTACTGCAGGCTAGTGAGTCGGAGTCTCACAGTGAGGTCAAGCACATTCCTGGTGGCGTTTATGCACTA **ATCTACCTCATAGGCTTTGATGGGACAACGGTAATCACCAGGGCTGTGGCCGCAAACAATGGGCTGACGA ACGAAAGAGTGGCAACAGGATCCGTCGTTACGGTCGCTGGGGTGAGCAACTTCGAGCTGATCCCAAATCC** CATTCCGGACGACACCCTGGAGAAGCACACTCTCAGGTCAGAGACCTCGACCTACAATTTGACTGTGGGG **GACACAGGGTCAGGGCTAATTGTCTTTTTCCCTGGATTCCCTGGCTCAATTGTGGGTGCTCACTACACAC** CTCCTCCTTCTACAACGCTATCATTGATGGTTAGTAGAGATCAGACAAACGATCGCAGCGATGACAAACC **666AGCCTAGCAGTGACGATCCATGGTGGCAACTATCCAGGGGCCCTCCGTCCCGTCACGCTAGTGGCCT** GGATACGATCGGTCTGACCCCGGG GGAGTCACCCGGGGACAGGCCGTCAAGGCCTTGTTCCAGGATGGGA **GCAAGATCAAACCCAACAGATTGTTCCGTTCATACGGAGCCTTCTGATGCCAACAACAGCGGCGGCGTC ATCCGGGCCATAAGGAGGATAGCTGTGCCGGTGGTCTCCACATTGTTCCCACCTGCCGCTCCCTAGCC CAGCTTACCCACATCATATGATCTTGGGTATGTGAGGCTTGGTGACCCCATTCCCGCAATAGGGCTTGA AATACTTCATGGAGGTGGCCGACCTCAACTCTCCCCTGAAGATTGCAGGAGCATTCGGCTTCAAAGACA** IGATGTCTGCAACAGCCAACATCAACGACAAAATTGGGAACGTCCTAGTAGGGGAAGGGGTCACCGTCC **AACGGCACCATAAACGCCGTGACCTTCCAAGGAAGCCTGAGTGAACTGACAGATGTTAGCTACAATGGG** ACCAATTCTCATCACAGTACCAACCAGGTGGGGTAACAATCACACTGTTCTCAGCCAACATTGATGCCA CACAAGCCTCAGCGTTGGGGGAGAGCTCGTGTTTCAAACAAGGGTCCACGGCCTTGTACTGGGCGCCACC **ATACTGAGTGAGGGGCCCTCTTGCCATCAAGACCGTCTGGCCAACAAGGGAGTACACTGACTTCGT** CCGCGTCAGGAAAAGCAAGAGCTGCCTCAGGCCGCATAAGGCAGCTGACTCTCGCCGCCGACAAGGGGT, **GAACTAGCAAAGAACCTGGTTACAGAATACGGCCGATTTGACCCAGGAGCCATGAACTACACAAAATT** CCAAAAATGGTAGCCACATGTGACAGCAGTGACAGGCCCAGAGTCTACACCATAACTGCAGCCGATGAT 8 ೩ 47 421 491 561 631 841 **=** 98 051 261 331 **40** 541 351 20 12 281 <u>6</u> <u>=</u>

Fig.5/

CAGATGAAAGATCTGCTCTTGACTGCGATGGAGGATGAAGCATCGCAATCCCAGGCGGGCTCTACCAAAGC **CCAAGCCAAAACCCAATGCTCCAACACAGAGACCCCCTGGTCGGCTGGGCCGCTGGATCAGGACCGTCTC** CACCTTGCCATGGCTGCATCAGAGTTCAAAGAGACCCCCGAACTCGAGAGTGCCGTCAGAGCAATGGAAG **GTGACTGACATGGCCAACTTCGCACTCAGCGACCCGAACGCCCATCGGATGCGAAATTTTCTTGCAAAC** SCACCACAAGCAGCAGCAAGTCGCAAAGGGCCAAGTACGGGACAGCAGGCTACGGAGTGGAGGCTCGGG **ACCCCAAGCTTTCATAGACGAAGTTGCCAAAGTCTATGAAATCAACCATGGACGTGGCCCAAACCAAGAA 16AAGACCTCCAACCTCCATCTCAAAGAGGATCCTTCATACGAACTCTCTGGACACAGAGTCTATGGA SGCGAGATTGAGAAAGTAAGCTTTAGAAGCACCAAAGCTCGCCACTGCACACCCGACTTGGCCTTAGGTTGG** CCACGCCACTGGGACAGGCTCCCCTACCTCAACCTACCATACCTTCCACCCAATGCAGGACGCCAGTAC **SCCCCACACCAGAGGAAGCACAGAGGGAAAAAGACACACGGATCTCAAAGAAGAGATGGAGCCATGGGCA**T ;TACTTTGCAACACCAGAATGGGTAGCACTCAATGGGCACCGAGGGCCAAGCCCCGGCCAGCTAAAGTAC **GATGAGGACCTTGAGTGAGGCTCCTGGGAGTCTCCCGACACCACCCGCGCGGGTGTGTGGACACCAATTCG** :TGGTCCCGGAGCATTCGATGTAAACACCGGGCCCAACTGGGCAACGTTCATCAAACGTTTCCCTCACAA **36GACGACAGCATTATGCTGTCCAAAGATCCCATACCTCCTATTGTGGGAAACAGTGGAAATCTAGCCAT ATGCTCCAGATGGGGTACTTCCACTGGAGACTGGGAGAGACTACACCGTTGTCCCAATAGATGATGTCT** AGCTTACATGGATGTTTCGACCCAAAGTCCCAATCCATGTGGCTATGACGGGAGCCCTCAATGCTTG1 CAGCAGCCAACGTGGACCCACTATTCCAATCTGCACTCAGTGTGTTCATGTGGCTGGAAGAGAATGGGA :GACAGTGGAAGACGCCATGACACCCAAAGCATTGAACAGCAAAATGTTTGCTGTCATTGAAGGCGTGC **SCCTTACAACATCCCAAATTGGATCCGTTCGCGGGTCCCCT** 2731 2801 2941 3011 3081 2241 2311 2381 2451 2591 2661 2871 2521 2031 2101 217

Total number of bases is: 3261.

DNA sequence composition: 873 A; 909 C; 847 G; 632 T; 0 OTHER;

Sequence name: D78F (SEQ ID NOS: 27 and 29)

Fig.5B

AGGA TCGTCGAGTGGATATTGGCCCCGGAAGAACCCAAGGCTCTTGTATATGCGGACAACATATACATTG aagg agagacaattggcgagatgatagctatctcaaaccagtttctcagagagctatcaacactgttgaa gcaaggtg cagggacaaaggggtcaaacaagaagctactcagcatgttaagtgactattggtactta FCAT GCGGGCTTTTGTTTCCAAAGGCTGAAAGGTACGACAAAGTACATGGCTCACCAAGACCCGGAACA AAAT AACGTGTTGAACATTGAAGGGTGTCCAT CACTCTACAAATTCAACCCGTTCAGAGGAGGGTTGAAC CCA CTCAAACACGTGGTACTCAATTGACCTAGAGAGGGTGAGGCAAACTGCACTCGCCAACACGTGCA **GCGAAGCACGATCTCAGCAGCGTTCGGCATAAAGCCTACTGCTGGACAAGACGTGGAAGAGCTCTTGATC** GCCACCTTTGCCATGAACATTGCCCCTGCTCTAGTGGTGGACTCATCGTGCCTGATAATGAACCTGCAAA CCTAAAGTTTGGGTGCCACCTGAGGATCCGCTTGCCAGCCCTAGTCGACTGGCAAAGTTCCTCAGAGAGA **IGAAGTAACCCTCTTGACCCAAAACATAAGGGACAAGGCCTATGGAAGTGGGACCTACATGGGACAAGCA ACACTTTTGAGAGCATCGCGCAGCTACTTGACATCACACTACCGGTAGGCCCACCCGGTGAGGATGACAA** SAGGTTGA AGATTACCTTCCCAAAATCAACCTCAAGTCATCAAGTGGACTACCATATGTAGGTCGCACCA TAAGACCTATGGTCAAGGCAGCGGGAATGCAGCCACGTTCATCAACAACCACCTCTTGAGCACACTAGT CTAGGTATCAACTTTAAGATTGAGAGGTCCATTGATGATATCAGGGGCAAGCTGAGACAGCTTGTCCTCC |TGCACAACCAGGGTACCTGAGTGGGGGGTTGAACCAGAACAATCCAGCCCAACTGTTGAGCTTGACCT SGATACGATGGGTCTGACCCTCTGGGAGTCACGAATTAACGTGGCTACTAGGGGCGATACCCGCCGCTGG CCCCACGTTAGTGGCTCCTCTTCTTGATGATTCTGCCACCATGAGTGACATTTTCAACAGTCCACGGC **ACATCGCACTACTCAAGCAGATGATTTACCTGTTTCTCCAGGTTCCAGAGGCCAACGAGGGCCTAAAGGA AATCGACTTGTGGCCATGAAGGAGGTCGCCACTGGAAGAACCCCAAACAAGGATCCTCTAAAGCTTGGGT** 8 50 841 631 98 421 491 561 **=** 051 261 331 **4**0 471 2 121 191 19

- ig.6/

TGTTGGGCTCCACCTGCCCGCCAAGAGAGCCACCGGTGTCCAGGCCGCTCTTCTCGGAGCAGGAACGAG CAGACCAATGGGGATGGAGGCCCCAACACGGTCCAAGAACGCCGTGAAAATGGCCAAACGGCGGCAACGC CAAAAGGAGGCCGCTAACAGCCATGATGGGAACCACTCAAGAAGAGGACACTAATCCCAGACCCGGTAT **3GAGAAAGCCGACATCGCCAGCAAGGTCGCCCACTCAGCACTCGTGGAAACAAGCGACGCCCTTGAAGCA** CTCGTCCTTCTAGCCACAGCAAGAAGCCGTCTGCAAGATGCAGTTAAGGCCAAGGCAGAAGCCGAGAAAC **GTTCAGTCGACTTCCGTGTACACCCCCAAGTACCCAGAAGTCAAGAACCCACAGACCGCCTCCAACCCCG** TTTGTTCT6CT6CGTATCCCAAGGGAGTAGAGAACAAGAGTCTCAAGTCCAAAGTCGGGATCGAGCAGG CATACAAGGTAGTCAGGTATGAGGCGTTGAGGTTGGTAGGTGGTTGGAACTACCCACTCCTGAACAAAGC CTGCAAGAATAACGCAGGCGCCGCTCGGCGCATCTGGAGGCCAAGGGGTTCCCACTCGACGAGTTCCTA CTGAGAGCCTAGCCGAACTGAACAAGCCAGTACCCCCCAAGCCCCCAAATGTCAACAGACCAGTCAACAC **ACTAGGGTGGTCAGCTACATACAGCAAAGATCTCGGGATCTATGTGCCGGTGCTTGACAAGGAACGCCTA** SCC6AGTGGTCTGAGCTGTCAGAGTTCGGTGAGGCCTTCGAAGGCTTCAATATCAAGCTGACCGTAACAT CCACAAGTCCAAGCCAGACGCCCGATGCAGACTGGTTCGAAAGATCAGAACTCTGTCAGACCTTC1 cccesccTTcsccTs csssscccc 2591 2381 2451 2521 2661 273 2241 203 231 196 210 2171

DNA sequence composition: 796 A; 770 C; 724 G; 537 T; 0 OTHER; Total number of bases is: 2827

Sequence name: P2B (SEQ ID No: 25)

Fig.6B

INTERNATIONAL SEARCH REPORT

International application No. PCT/US97/12955

A. CLA	SSIFICATION OF SUBJECT MATTER			
	:Please See Extra Sheet. :Please See Extra Sheet.			
According	to International Patent Classification (IPC) or to both	h national classification and IPC		
B. FIEI	LDS SEARCHED			
Minimum d	documentation searched (classification system follow-	ed by classification symbols)		
U.S. :	424/184.1, 204.1, 816, 826; 435/71.1, 235.1, 236,	237, 238, 239, 320.1; 536/23.72		
Documenta	tion searched other than minimum documentation to th	e extent that such documents are included	in the fields searched	
Electronic	data base consulted during the international search (n	ame of data base and, where practicable	e, search terms used)	
i .	N-MEDLINE, BIOSIS, CAPLUS, CABA	*		
C. DOC	UMENTS CONSIDERED TO BE RELEVANT			
Category*	Citation of document, with indication, where a	ppropriate, of the relevant passages	Relevant to claim No.	
Х	MUNDT et al. Complete Nucleotid Noncoding Regions of Both Genome S of Infectious Bursal Disease Virus. Vir 10-18, see entire document.	Segments of Different Strains	1-2, 4-20	
X	US 4,530,831 A (LUTTICKEN ET A) see entire document.	L) 23 JULY 1985 (07/23/85),	7, 15-20	
x	US 5,192,539 A (VAN DER MARE (09/03/93), see entire document.	L ET AL) 09 MARCH 1993	1-3, 7, 15-20	
х	MUNDT et al. Identification of a not bursal disease virus-infected cells. Je 1995, Vol. 76, pages 437-443, see en	ournal of General Virology.	8	
			·	
X Furt	her documents are listed in the continuation of Box (C. See patent family annex.		
Special entagories of cited documents: "T" later document published after the international filling date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention				
to be of particular relevance "E" earlier document published on or after the international filing date "X" document considered		considered novel or cannot be conside		
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) *O* document referring to an oral disclosure, use, exhibition or other combined with one or more other such interest.		step when the document is a document, such combination		
•P• do	coment published prior to the international filing data but later than s priority data claimed	"A." document member of the same patent		
	actual completion of the international search	Date of mailing of the international sec	arch report	
22 SEPT	EMBER 1997	1 0 NOV 1997		
22 SEPTEMBER 1997 Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-3230 Telephone No. (763) 308-9196				

INTERNATIONAL SEARCH REPORT

International application No. PCT/US97/12955

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No
K	BAYLISS et al. A comparison of the sequences of segment A of four infectious bursal disease virus strain and identification of a variable region in VP2. Journal of General Virology. 1990, Vol. 71, pages 1303-1312, see entire document.	1-2, 5-8, 10-13
<i>t</i>	MORGAN et al. Sequence of the Small Double-Stranded RNA Genomic Segment of Infectious Bursal Disease Virus and Its Deduced 90kDa Product. Virology. 1988, Vol. 163, pages 240-242, see entire document.	1-20
,	SPIES et al. Nucleotide sequence of infectious bursal disease virus genome segment A delineates two major open reading frames. Nucleic Acids Research. 1989, Vol. 17, No. 19, page 7982, see entire document.	1-20
	WO 91/16925 A1 (UNIVERSITY OF MARYLAND at COLLEGE PARK) 14 NOVEMBER 1991 (14/11/91), see entire document.	1-20
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INTERNATIONAL SEARCH REPORT

International application No. PCT/US97/12955

A.	CLASSIFICATION OF SUBJECT MATTER:	
IP	C (6):	

A61K 39/00, 39/38, 39/12; C12P 21/04; C12N 7/00, 7/01, 7/02, 7/04, 7/06, 7/08, 15/00, 15/09, 15/63, 15/70, 15/74

A. CLASSIFICATION OF SUBJECT MATTER: US CL :

424/184.1, 204.1, 816, 826; 435/71.1, 235.1, 236, 237, 238, 239, 320.1; 536/23.72